ROLE OF ALLERGIC SENSITIZATION, FILAGGRIN VARIANTS, AND DNA

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ROLE OF ALLERGIC SENSITIZATION, FILAGGRIN VARIANTS, AND DNA METHYLATION ON THE RISK OF ALLERGIC DISORDERS

by

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DEDICATION

I dedicate this work to my parents who have provided me with unconditional support throughout my life and who inspired and encouraged me to pursue my educational ambitions. To my two sisters whose support fortified my educational success. A special feeling to my little sister, Fatima, being a Down syndrome girl has always made you so special and you will always have a special place in my heart.

I also dedicate this dissertation to my beloved wife who ensured my success through ongoing support, patience, and encouragement.

To my beloved daughter, Retaj (4 years old), who has been an unmatched source of inspirations during the course of my studies.

To my mentor, Prof. Wilfried Karmaus, who supported me and my family in exceptional manners. Prof. Karmaus, set the bar high from day one and guided me to that bar.
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ABSTRACT

Background: Allergic disorders, including eczema, asthma, and rhinitis, have emerged as a global public health concern due to their elevated prevalence and the associated clinical morbidity. Environmental, immunologic, and genetic factors have been implicated in the pathogenesis of allergic disorders. Allergic sensitization (representing deviated immune responses) and filaggrin gene (FLG) variants (leading to dysfunctional epidermal barrier) have shown to be common predisposing factors in the development of allergic disorders. However, there is a lack of knowledge on their joint effects on the development of single and multiple (coexistence) allergic disorders. More recently, epigenetic mechanisms, such as DNA methylation, have emerged as potentially important factors in the development of such complex diseases; however, the extent to which DNA methylation associates with allergic disorders is unclear.

Objectives: This dissertation sought to (i) determine whether eczema and/or allergic sensitization is an effect modifier of the association between ‘FLG variants and asthma’ and ‘FLG variants and rhinitis’, (ii) test whether FLG variants and allergic sensitization jointly predispose to the comorbidity of eczema, asthma and rhinitis, and (iii) examine associations between DNA methylation across the epidermal differentiation complex (EDC) genomic region with eczema status.

Methods: The Isle of Wight (IOW) birth cohort, a population-based sample of 1,456 infants born between January 1989 and February 1990, was prospectively assessed at ages 1, 2, 4, 10, and 18 years. Repeated measurements of eczema, asthma, rhinitis, and
allergic sensitization (documented by skin prick tests) were available for all follow-ups.

*FLG* variants R501X, 2282del4, and S3247X were genotyped in 1,150 participants. Log-binomial regression models were applied to test for associations and statistical interactions on multiplicative scale. On the other hand, DNA methylation was measured in a subsample (n = 367) of the IOW participants at age 18 years (*discovery cohort*) and in two semi-independent samples (*replication cohorts I and II*). Associations between eczema status and DNA methylation were assessed using linear regression.

**Results:** *FLG* variants were associated with increased risk of asthma and rhinitis. Both eczema status (RR<sub>interaction</sub> = 1.96, *P*<sub>interaction</sub> = 0.006) and allergic sensitization (RR<sub>interaction</sub> = 1.58, *P*<sub>interaction</sub> = 0.013) modified the association between *FLG* variants and asthma, but not the association with rhinitis. The combined effect of both risk factors increased the risk of coexisting “eczema and asthma” (RR = 13.67, 95% CI: 7.35 – 25.42), “asthma and rhinitis” (RR = 7.46, 95% CI: 5.07 – 10.98), and “eczema, asthma, and rhinitis” (RR = 23.44, 95% CI: 12.27 – 44.78). On the other hand, Differential DNA methylation of CpG site cg12048339 (located within promoter of *S100A6* gene) was associated with eczema specifically among female participants of all study cohorts; whereas, aberrant DNA methylation of cg10959711 (located within promoter of *S100A11* gene) associated with eczema among male participants in all study samples.

**Conclusions:** Allergic sensitization and eczema modulated the association between *FLG* variants and asthma, but not rhinitis; implying that the mechanisms and pathways through which *FLG* variants predispose to increased risk of asthma and rhinitis may be different. Moreover, the coexistence of allergic disorders is frequent and allergic sensitization and *FLG* variants jointly increased risk of allergic comorbidities, which may represent more
severe and complex clinical phenotypes. Results of an exploratory investigation demonstrated that DNA methylation of the EDC locus could be an important factor in the development of eczema in a sex-specific manner. Future studies corroborating our findings are needed.
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LIST OF ABBREVIATIONS

CI.......................................................................................................................... Confidence Interval
CpG.................................................................................................................... Cytosine-Guanine dinucleotides
DNA................................................................................................................. Deoxyribonucleic acid
EDC.......................................................................................... Epidermal Differentiation Complex
FLG................................................................................................................... Filaggrin gene
GEE........................................................................................................... Generalized Estimating Equations
IgE................................................................................................................... Immunoglobulin E
IOW...................................................................................................................... Isle of Wight
ISAAC........................................................International Study of Asthma and Allergies in Childhood
PAF.............................................................................................................. Population Attributable Fraction
RR................................................................................................................ Risk Ratio
SA.................................................................................................................. Specific Aim
SPT............................................................................................................... Skin Prick Test
CHAPTER 1
INTRODUCTION

1.1 BACKGROUND

Allergic disorders, including eczema, asthma, and rhinitis, have emerged as a global public health concern [1]. The clinical morbidity associated with allergic disorders that leads to social and economic burden on families and societies further classify these disorders as major global public-health challenges [1, 2]. The direct annual cost associated with clinical management of eczema alone was estimated to be £47 ($76) million in the United Kingdom, with families bearing £17 ($27) million of disease costs [3]. Furthermore, Asthma has even greater economic burden, costing £843 million ($1.368 billion) annually [3]. The elevated economic burden and clinical morbidity associated with such chronic diseases, clearly demonstrates the need for developing cost-effective intervention and prevention strategies that aim at reducing the burden associated with allergic disorders. However, to reach the aforementioned goal, there is a need to better understand the natural history and the biological mechanisms that underlie the clinical manifestation of these complex disorders at different developmental stages.

The International Study of Asthma and Allergies in Childhood (ISAAC) provides unmatched opportunity to explore the trends in prevalence of allergic disorders over time [4]. Reports based on the ISAAC study revealed the existence of global variations in the prevalence of allergic disorders [5-8]. During the last 3 decades, industrialized nations witnessed disproportionally increased trends in the prevalence of eczema, asthma, and
However, recent reports are indicating that a plateau in the prevalence has been reached in most of the countries that experienced increasing trends [10-12]. In contrast, countries with formerly low-prevalence are reporting increasing patterns in the prevalence and incidence of allergic disorders [10-13]. The disparity in the prevalence across nations and the changing trends over time within populations are indicators of the importance of environmental factors, in addition to the genetic elements, in disease pathogenesis. Deeper understanding of the natural history and risk factors of these chronic conditions will help us elucidate their plausible underlying etiological mechanisms.

1.2 OVERALL STRUCTURE OF THE DISSERTATION

This document is divided into six chapters, as the following:

i. The first chapter provides background information on the dissertation topic, literature review relevant to the study questions, current gap in knowledge, and the specific aims and hypotheses of this study.

ii. The second chapter explains in details the methodological parts of the study and the statistical analyses that were applied to achieve the goals of this dissertation.

iii. Each of chapter three, four, and five is organized and formatted as a journal research report that have been or will be submitted for publication. Each of these three research-related chapters, also referred to as results chapters, represent one of the three specific aims of this dissertation.
iv. The sixth, last, chapter provides an overall synopsis and discussion of the results of the three research-related chapters and how results of these chapters are synthesized.

1.3 LITERATURE REVIEW

1.3.1 GLOBAL VARIATIONS IN THE PREVALENCE OF ALLERGIC DISORDERS

Reports based on the ISAAC study revealed the existence of global variations and both between and within countries variations in the prevalence of allergic disorders [5, 10, 11, 14, 15]. For instance, for the age group 6 to 7 years, data based on the ISAAC study showed that the 12-month prevalence of eczema ranged from 0.9% in India to 22.5% in Ecuador [14]. For the age group 13 to 14 years, the prevalence of eczema between countries ranged from 0.2% in China to 24.6% in Columbia [14]. Similarly, the prevalence of asthma symptoms among 13 to 14 years old children ranged from 0.1% in Pune (India) to 16% in Costa Rica [16]. Prevalence of rhinitis showed even wider global variation, which ranged from 1% in India to 45.1% in Paraguay [17]. In addition to estimating the prevalence and time trends of allergic disorders, the ISAAC study allowed researcher to discover wide-range of geographical differences in the prevalence of eczema within countries. For example, the ISAAC phase 3 study was conducted in 10 different locations across Spain, with the lifetime prevalence among 6 to 7 years olds being 35.6% in ‘A Coruna’ and 25.0% in ‘Barcelona’ [14]. Although the role of environmental factors in explaining the between and within countries variations in allergic disorders prevalence is essential, there is some strong evidence suggesting that genetic predisposition can potentiate the effects of environmental factors [18]. In
summary, both the *between* and *within* countries differences in the prevalence of allergic disorders are worth remarking as they can contribute to the advancement of etiologic research.

1.3.2 Natural History of Allergic Disorders

Eczema is an inflammatory skin disorder that is characterized by disrupted epidermal barrier function, immunoglobulin E (IgE)-mediated sensitization to food and environmental allergens (i.e., allergic sensitization), and a remitting–relapsing combination of skin dryness and itching [19-21]. Globally, the prevalence of eczema among children and adults has been estimated to be between 15% to 30% and 2% to 10%, respectively, demonstrating wide variation across nations [20, 22]. The three major developmental periods of eczema are: infancy, childhood, and adolescence/adulthood; with 45% of all eczema cases occurring in the first 6 months of life [20, 23]. However, the long-term prognosis (natural history) of eczema is complex due to the relapsing and remitting nature of the disease. Emerging evidence is suggesting that eczema affects more boys during early life stages and becomes more predominant among adolescent girls and thereafter [20, 24]. Recently, we demonstrated that girls develop more eczema during puberty period while more boys outgrow their eczema [25]. Yet, more insights on the etiology of eczema are needed in order for preventable public-health efforts to succeed.

Likewise, population-based studies have shown that the prevalence of both asthma and rhinitis is increasing and disproportionally affecting developed and developing countries [10]. Asthma, accounting for a substantial proportion of
burden/morbidity associated with allergic disorders, is a common chronic disorder affecting up to 25% of children and adults around the world [16, 26-29]. The major pathophysiological hallmarks that contribute to the complex heterogeneity of the clinical manifestations of asthma are recurrent airway inflammation, variable airway obstruction, and bronchial hyper-responsiveness [30, 31]. Similarly, an increase in the prevalence of rhinitis (10% to 30%), which is an inflammatory disease affecting the nasal tissue, has been documented during the last 30 years [32]. Common symptoms of rhinitis are nasal congestion and itching, sneezing, and rhinorrhea, which negatively impact the quality of life [32, 33]. The natural history of both asthma and rhinitis is influenced by gender [34, 35]. For asthma, a gender reversal occurs around the puberty period; the disorder predominantly affects boys before puberty, whereas, more girls are affected during adolescence and in early adulthood [26, 35-37]. However, the effect of gender on the long-term prognosis of rhinitis is further complicated by allergic sensitization (defined later) [33, 34].

Although it is acceptable to investigate the etiological mechanisms underlying allergic diseases separately; evidence supporting their comorbidity is increasing [38, 39]. Allergic comorbidity (coexistence) is defined as the concurrent presence of more than one allergic disorder (i.e. eczema, asthma, and rhinitis) in the same individual. The elevated comorbidity of asthma (lower-airway disorder) and rhinitis (upper-airways disorder) and their shared pathophysiology further support the concept of “one airway one disease” [12, 40]. In addition to the comorbidity of the allergic respiratory diseases, the concept of “allergic march” suggests that allergic diseases develop in a sequential pattern. Eczema is considered as the first step in the allergic march, usually developing
early in infancy/childhood followed by manifestations of allergic disorders of the airway later in childhood or early adolescence period [41]. The continuum developmental mechanism of allergic diseases is widely discussed in the scientific literature; however, supporting replicable evidence and high consensus is far from complete [41, 42]. Another revealing concept is the “coexistence” of allergic morbidities, which opposes the linear developmental pathway [42-44]. A report based on the German Multicenter Atopy Study (MAS), showed that eczema alone early in life does not support the allergic march, rather the coexistence of eczema and wheezing predicted asthma [45]. Such observation speaks in favor of the coexistence of allergic morbidities instead of a progressive development. The interrelationship between these conditions and the risk factors that either predispose individuals to follow the “allergic march” or “coexistence” remain an open field for research.

1.3.3 Allergic Sensitization and Filaggrin Variants: as Common Risk Factors

A complex interplay between genetic, environmental, and immunological factors is considered to contribute to the pathogenesis of allergic disorders [43, 46]. Allergic sensitization, defined as the genetic susceptibility to produce Immunoglobulin E (IgE) antibodies in response to exposure to environmental or food antigens, is the most common thread linking the manifestations of allergic disorders [46-48]. Arshad et al suggested that 30% to 40% of cases of eczema, asthma, and rhinitis in early childhood can be attributed to allergic sensitization [49]. Yet, others suggested that up to 80% of asthmatic patients have concomitants allergic sensitization [50]. Similarly, it is well established that manifestations of eczema and rhinitis are closely associated with allergic
sensitization [23, 32, 46, 48]. Candidate-gene and genome-wide association studies have identified several genetic variants that are associated with the observed variability in IgE levels between allergic and non-allergic individuals [51, 52]. As a result, genetic factors that regulate the immune response dominated the research of allergic diseases for the last 30 to 40 years.

The recent discovery of loss-of-function variants in the filaggrin gene (FLG) caused a shift in the research paradigm from mainly focusing on immune related genes to incorporating genetic factors that regulate the formation of the epidermal barrier [53-55]. Filaggrin (filament-aggregating protein) is a key protein for the formation of functional skin barrier that inhibits the penetration of allergens, microbes, and irritants and limits transepidermal water loss [55-57]. Loss-of-function variants within the FLG are the strongest and most replicated risk factors for eczema development, yet [58, 59]. Filaggrin haploinsufficiency, i.e. reduction in filaggrin protein expression in heterozygous individuals, is associated with approximately 3-fold increased risk for eczema [60, 61]. It has been estimated that around 25% to 50% of eczema patients are carriers of at least one FLG variant, which is associated with an estimated population-attributable risk ranging from 4.2% to 15.2% [58, 61-63]. In addition to its association with eczema, multiple studies showed that FLG variants are associated with asthma and rhinitis [56, 60]. One school of thought considers that the association between FLG variants and asthma depends on the presence of eczema [59, 64, 65]. However, there is conflicting evidence as to whether the association between FLG variants and rhinitis is modified by eczema status [63, 64, 66]. To this end, it is clear that FLG variants are significant common thread for the development of the different allergic disorders.
1.3.4 DNA Methylation and the Risk of Eczema

The high heritability of eczema (71% to 84%), asthma (35% to 95%), and rhinitis (33% to 91%) and clustering within families demonstrate the importance of genetic predisposition in their pathogenesis [43, 67, 68]. However, thus far candidate-gene and genome-wide association studies have failed to explain the observed high heritability of allergic diseases. Hence, suggesting that genetic regulatory factors, other than DNA sequence variants, such as epigenetic variants may account for the unexplained genetic effect [69]. Epigenetic regulatory mechanisms are mitotically heritable and can alter gene activity without changing the DNA sequence [70]. DNA methylation, widely studied in epidemiological investigations due to practical and biological reasons [71, 72], along with histone modifications are important epigenetic marks that work hand-in-hand on influencing disease expression [73]. Since eczema is considered as the first manifestation of allergic disorders that can predispose to subsequent allergic morbidities, it is thus important to understand the epigenetic etiologic mechanisms that underlie eczema development. Up to our knowledge, the impact of DNA methylation on the risk of eczema has not been investigated previously. The epidermal differentiation complex (EDC), located on human chromosome 1q21, harbors a dense cluster of genes (including FLG) that are involved in the terminal differentiation of keratinocytes that are responsible for the integrity and functionality of the epidermal barrier [74-76]. Previous reports showed that a compromised epidermal barrier predispose to the development of eczema [77, 78]. Therefore, a comprehensive DNA methylation profiling of the EDC genomic region should provide novel insights into the epigenetic contribution in the development of eczema.
1.4 Gaps in Current State of Knowledge

Both, allergic sensitization (deviated immune response) and FLG variants (defective epidermal barrier) are important risk factors that link and predispose to the development of allergic disorders[46]. However, the current state of knowledge lacks clear understanding on the joint contribution of these two risk factors. Prior studies focused on eczema as the effect modifier for the association between FLG variants and asthma, while ignoring the role of allergic sensitization. Similarly, eczema was investigated as an effect modifier for the association between FLG variants and rhinitis, but not allergic sensitization. Furthermore, the single and joint role of FLG variants and allergic sensitization on the coexistence of allergic diseases (i.e., eczema, asthma, and rhinitis) has not been previously investigated. On the other hand, no existing knowledge on the role of epigenetic regulatory mechanisms (e.g., DNA methylation) of the EDC genomic loci that controls the epidermal barrier formation and functionality in relation to eczema risk is available.

1.5 Study Proposal, Hypotheses, and Aims

Population-based longitudinal studies provide an unmatched opportunity to investigate the natural history of allergic disorders. Cross-sectional and case-control studies lack the ability of determining the longitudinal effect of risk factors on outcomes. However, follow-up studies with repeated measurements of the outcomes of interest (e.g., eczema, asthma, and rhinitis) and risk factors (e.g., allergic sensitization) facilitate the determination of whether the effects are established early or later in the development of diseases. Furthermore, we can determine whether the effect of time-independent risk
factor (e.g. FLG variants) changes over-time. In contrast, for dynamic (time-varying) risk factors (e.g., allergic sensitization) determining their concurrent effects allow the risk to change over-time. This dissertation aims at enhancing our understanding of the role of FLG variants, allergic sensitization, and epigenetic modifications on the development of allergic diseases, by examining the following specific hypotheses (H):

**H1:** Allergic sensitization, not eczema, is the main effect modifier of the association between FLG variants and asthma.

**H2:** The association between FLG variants and rhinitis is modified by allergic sensitization status, not eczema status.

**H3:** FLG variants and allergic sensitization jointly increase the risk of having single and multiple (coexisting) allergic disorders.

**H4:** Eczema-affected and eczema-free individuals have different DNA methylation profiles for the EDC genomic region.

Hypothesis 1, 2, and 3 will be tested using data from the Isle of Wight 1989 birth cohort study (n = 1,456). Participants were prospectively followed-up at the ages of 1, 2, 4, 10, and 18 years. Hypothesis 4 will be tested in a subset (n = 367) of Isle of Wight birth cohort participants that had DNA methylation measurements. For the purpose of validation, hypothesis 4 will be further tested in two semi-independent samples (pregnant cohort participant, their partners, and their offspring, the F2 generation). The specific aims (SA) of this dissertation are:

**SA1:** To determine the magnitude of modification imposed by eczema and allergic sensitization on the association between “FLG variants and asthma” (H1).

Similarly, I will determine whether eczema and/or allergic sensitization act as an
effect modifier for the association between “FLG variants and rhinitis” (H2).

Overall, I will test whether the combined (joint) effect of two risk factors (i.e., “FLG variants and eczema” or “FLG variants and allergic sensitization”) yields higher risk of asthma (or rhinitis).

SA2: To test whether FLG variants and allergic sensitization jointly predispose to the comorbidity of eczema, asthma and rhinitis (H3).

SA3: To associate DNA methylation across the EDC genomic region with eczema status (H4).

1.6 How Would Results of This Dissertation Enhance Our Knowledge?

Better understanding of the genetic and epigenetic contribution to the development of allergic diseases will be gained by successfully accomplishing the specific aims of this dissertation. We have previously shown that the combined effect of allergic sensitization and FLG variants predisposed individuals at higher risk of having concurrent and subsequent eczema [79]. In this dissertation, I will investigate whether allergic sensitization (or eczema) in combination with FLG variants increase the risk of having concurrent and subsequent asthma and rhinitis. The current knowledge indicates that FLG variants increase the risk of asthma only in the presence of eczema [59, 66]; however, I propose to test whether allergic sensitization is the effect modifier rather than eczema. Furthermore, the current inconsistencies in the associations between FLG variants and rhinitis can be attributed to the failure of previous studies in accounting for the possible interaction between FLG variants and allergic sensitization on the risk of rhinitis. Therefore, SA1 of this dissertation will enhance our knowledge by showing that
allergic sensitization is the main effect modifier of the association between FLG variants and asthma and FLG variants and rhinitis.

Results of SA1 of this dissertation will show the importance of FLG variants and allergic sensitization on the development of single allergic disorders (i.e., asthma and rhinitis). In contrast, SA2 sought to determine whether FLG variants and allergic sensitization are associated with the coexistence of multiple allergic diseases rather than a single morbidity. Previous etiologic research in the field of allergic comorbidities is highly limited and assessments of the joint role of allergic sensitization and FLG variants on the development of allergic comorbidities is lacking. Therefore, results of SA2 will enhance our understanding of the underlying risk factors that predispose individuals at higher risk of having multiple allergic disorders.

In addition to the role of genetic variants in the pathogenesis of allergic diseases, the etiologic role of epigenetic modifications (e.g., DNA methylation) is rapidly emerging and scientifically sound. Recently we showed a significant interaction between FLG variants and adjacent differential DNA methylation on the risk of eczema [80]. Results of this report suggested that the association between FLG variants and eczema is modulated by DNA methylation. Recent evidence is suggesting that epigenetic mechanisms, such as DNA methylation, that alter gene activity without changing the underlying DNA sequence account for a considerable amount of the unexplained genetic effect found in complex diseases, such as eczema [69, 72]. In this dissertation I propose going beyond the single-gene approach (i.e., FLG gene) into incorporating the DNA methylation of the EDC locus at chromosome 1q21 (SA3). The EDC includes a dense cluster of approximately 60 genes (including FLG) encoding structural and regulatory
proteins that are essential for keratinocyte differentiation [76, 81, 82]. Comprehensively analyzing DNA methylation status of the EDC genomic region will provide novel insights, beyond genetic variants, on the contribution of epigenetic modifications. Therefore, I anticipate that DNA methylation of the EDC will further show the importance of this genomic region for the development of eczema. SA3 will focus on eczema since it is considered the first manifestation of allergic diseases. The ultimate goal of this dissertation is to improve our ability of characterizing and classifying susceptible individuals, which will lead to an improved clinical management and the development of efficient intervention and prevention strategies.
CHAPTER 2

METHODS

2.1 STUDY DESIGN AND PARTICIPANTS

This dissertation is mainly based on information collected from the Isle of Wight (IOW) birth cohort. An unselected population-based study (n = 1,536) was recruited between January 1989 and February 1990 in the Isle of Wight, United Kingdom, to prospectively study the natural history and etiology of asthma and allergic diseases. The island is close to the British mainland, semi-rural, without heavy industry. Both the Isle of Wight and the study populations are 99% Caucasian. After excluding adoptions, perinatal deaths, and refusals for follow-up, written informed consent was obtained from parents to enrol 1,456 children (95%), with follow-up assessments conducted at ages 1 (n = 1,167), 2 (n = 1,174), 4 (n = 1,218), 10 (n = 1,373), and 18 (n = 1,313) years. Ethics approvals were obtained from the Isle of Wight Local Research Ethics Committee (now named the National Research Ethics Service, NRES Committee South Central – Southampton B) at recruitment and for the 1, 2, 4, 10 and 18 years follow-ups (06/Q1701/34). Detailed questionnaires were completed for each child at each follow-up. When a visit was not possible, a telephone questionnaire was completed or a postal questionnaire sent for completion and return.

For the purpose of testing H4, a subsample (discovery cohort: n = 367, aged 18 years) of the IOW birth cohort participants were randomly selected for epigenetic study. We also considered two cohorts to test the reproducibility of results obtained from the
discovery cohort. **Replication cohort I** (n = 146; parents of the F2 generation) includes participants of the IOW cohort (28.8% were part of the discovery cohort) plus new participants. Expecting mothers and their partners were assessed before delivery. Replication cohort II (n = 94; F2 generation) includes the F2 newborns of the IOW cohort participants plus new parents. Infants were followed-up at ages 3-, 6-, and 12-months.

2.2 **Phenotypes**

In all assessments of the Isle of Wight birth cohort, eczema was defined as chronic or chronically relapsing, itchy dermatitis lasting more than 6 weeks with characteristic morphology and distribution [83], following Hanifin and Rajka criteria [84]. For asthma, at the 1, 2, and 4-year follow-ups, the medical investigator determined the presence of asthma based on wheeze frequency over the last 12 months and treatment given for asthma or asthma related symptoms. At the 10 and 18 year follow-ups, asthma was defined as having “ever had asthma” and either “wheezing or whistling in the chest in the last 12 months” or “current treatment for asthma”, using ISAAC questionnaire [4]. Rhinitis was defined by a positive response to: ‘In the past 12 months have you had a problem with sneezing, or a runny or a blocked nose when you did not have a cold or the flu?’ [33]. Since the 1-year and 2-year follow-up data on eczema, asthma, and rhinitis were collected in a relatively small time window, we combined them for analytic purposes (reported as 1-or-2 years).

To determine allergic sensitization status, skin prick testing (SPT) at ages 1 and 2 years was performed on children with any symptoms of eczema, asthma, or rhinitis. We combined SPT results for ages 1 and 2 years, since they occurred within a short time
period and will henceforth refer to this as SPT at 1-or-2 years. At 4, 10 and 18 years, regardless of symptoms, SPT was performed on most children attending the research center to a standard battery of common allergens (ALK-Albello, Horsholm, Denmark). Inhalant allergens tested were house dust mite, cat, dog, *Alternaria alternata*, *Cladosporium herbarium*, grass pollen mix, and tree pollen mix. Food allergens tested were cows’ milk, soya, hens’ egg, peanut and cod. Positive and negative controls were included. Allergic sensitization was defined by having a SPT to at least one allergen test with mean wheal diameter of 3 mm greater than the negative control. Since allergic sensitization is a dynamic rather than a completely stable phenotype, we used the concurrent status and thus allowed the risk to change over time.

2.3 *FLG* GENOTYPING

DNA was extracted from blood or saliva samples from cohort subjects (n = 1,211). Five variants in the *FLG* gene that result in loss of function and are reported to be common in populations of European ancestry were selected for genotyping [79]. DNA samples were interrogated using GoldenGate Genotyping Assays (Illumina, Inc, SanDiego, CA) on the BeadXpressVeracode platform (Illumina, Inc, SanDiego, CA) per Illumina’s protocol. In brief, samples were fragmented and hybridized to the pool of allele-specific primer sets. Following an extension/ligation reaction the samples were then hybridized to the Veracode bead pool and processed on the BeadXpress reader. Data were analyzed using the genotyping module of the GenomeStudio Software package (Illumina, Inc, SanDiego, CA). DNA from each subject plus 37 replicate samples were analyzed for a total of 1,248 samples. The quality threshold for allele determination was
set at a GenCall score > 0.25 (scores ≤ 0.25 were “no calls”) with n = 1,227 samples (98.3%) retained for further analysis. Analysis of each locus included reclustering of genotyping data using our project data to define genotype cluster positions with additional manual reclustering to maximize both cluster separation and the 50th percentile of the distribution of the GenCall scores across all genotypes (50% GC score).

Children were classified as having \textit{FLG} loss-of-function defect if they carry the minor allele for at least one of the following \textit{FLG} null variants: R501X, 2282del, or S3247X. Variants 3702delG and R2447X were not informative in our population due to minor allele frequencies being < 0.1% [79], and thus were not used in defining filaggrin haploinsufficiency.

2.4 DNA Methylation Profiling

DNA was extracted from whole blood collected at age 18 years (\textit{discovery cohort}), whole blood of \textit{replication cohort I} participants’, and cord blood of \textit{replication cohort II} participants’ [85], and bisulfite-treated for cytosine to thymine conversion using the EZ 96-DNA methylation kit (Zymo Research, CA, USA). Genome-wide DNA methylation was assessed using the Illumina Infinium HumanMethylation450 BeadChip (Illumina, Inc., CA, USA), which interrogates > 484,000 CpG sites associated with approximately 24,000 genes. Arrays were processed using a standard protocol [86]. The BeadChips were scanned using a BeadStation, and the methylation level (beta value) calculated for each queried CpG locus using the Methylation Module of BeadStudio software. After cleaning of the DNA methylation data, beta (\(\beta\)) values presented as the proportion of methylated (M) over methylated (M) and unmethylated (U) sites (\(\beta=M/[c+M+U]\), with c
being constant to prevent dividing by zero) were used to estimate the effect of DNA methylation [87].

2.5 Statistical Analysis

Statistical analyses and approach for each of the three specific aims will be presented separately.

2.5.1 Specific Aim 1: Statistical Analysis

**SA1:** To determine the magnitude of modification imposed by eczema and allergic sensitization on the association between “FLG variants and asthma” (H1).

Similarly, I will determine whether eczema and/or allergic sensitization act as an effect modifier for the association between “FLG variants and rhinitis” (H2).

Overall, I will test whether the combined (joint) effect of two risk factors (i.e., “FLG variants and eczema” or “FLG variants and allergic sensitization”) yields higher risk of asthma (or rhinitis).

For all of the association analyses that will be performed to approach SA1, log-binomial (log linear) regression models will be applied to estimate risk ratios (RRs) and their 95% confidence intervals (95% CIs) using the GENMOD procedure in SAS 9.3 (SAS, Gary, NC, USA). Since the proposed effect modifiers (i.e., allergic sensitization and eczema) and the two outcomes (i.e., asthma and rhinitis) were measured repeatedly at ages 1-or-2, 4, 10, and 18 years, the generalized estimating equations (GEE) method will be used, which accounts for the correlated observations and the within-child effect by employing a covariance matrix solved through an iterative estimating process based on a
working correlation matrix [88]. The appropriate working correlation matrix will be determined by using the Akaike information criterion (AIC).

In an explanatory step, to determine whether the RRs of the association of FLG variants with asthma and rhinitis differ according to the presence or absences of the proposed effect modifiers, we evaluated the association in the total study sample and sub-samples based on the status of the possible effect modifiers. Next, interaction terms, on a multiplicative scale, were used to test the additional effect of two co-occurring risk factors on the health outcomes above and beyond their individual effects. Also, to evaluate whether the effect of FLG variants across levels of the effect modifiers were statistically significantly different, we included interaction terms in separate regression models (model 1: FLG variants × eczema and model 2: FLG variants × allergic sensitization). To decipher which interactive effect was more pronounced, a regression model including both interaction terms (model 3: FLG variants × eczema and FLG variants × allergic sensitization) was evaluated. Henceforth, we refer to models 1, 2, and 3 as the ‘concurrent models’ since the effect modifiers and the outcomes coexisted at the same time. In the case of a possibly statistically significant interaction term ($P_{interaction} < 0.1$), the “combined effect”, referring to the joint impact of two individual risk factors plus their interaction on the occurrence of the outcome, was estimated as follows: $RR = \exp[(\beta_1 \times \text{allergic sensitization}) + (\beta_2 \times FLG \text{ variants}) + (\beta_3 \times \text{allergic sensitization} \times FLG \text{ variants})]$. We repeated the previous analyses while stratifying by sex to determine if sex-specific effects existed. In all GEE models, sex and age at follow-up were included as potential confounders.
In addition to the concurrent models, we tested the interactions in ‘delayed effect’ models to investigate whether the time-order of risk and response supports the concurrent model findings. The delayed effect analysis helps to determine whether the interaction effects contribute to the developmental process of asthma and rhinitis. To this end, we analyzed whether FLG variants interact with preceding allergic sensitization or eczema on the risk of subsequent asthma and rhinitis. For example, the interaction of FLG variants with allergic sensitization status at age 1-or-2 years on the risk of asthma at age 4 years (FLG variants × SPT results at 1-or-2 years → asthma at 4 years) was evaluated. The prevalence of the clinical phenotype (e.g., asthma at 4), which was used in the delayed effect models, included new occurrences and persistence of the disease. Therefore, we constructed three delayed periods (1-or-2 to 4 years, 4 to 10 years, and 10 to 18 years) to determine if the presence of both FLG variants and preceding allergic sensitization or preceding eczema influences the positive transition, defined as the change in disease status from disease-free to diseased, and the persistence of the disease in two consecutive follow-ups.

2.5.2 Specific Aim 2: Statistical Analysis

**SA2:** To test whether FLG variants and allergic sensitization jointly predispose to the comorbidity of eczema, asthma and rhinitis (H3).

Information on eczema, asthma, and rhinitis were collected prospectively at ages 1-or-2, 4, 10, and 18 years from the IOW study participants. For each age I will create a new variable that has 8 categories, which identifies all the possible combinations of allergic comorbidities at the respective follow-up (Table 2.1). This type of categorization
will allow us to have better understanding on the prevalence of each allergic category at
the different ages.

Table 2.1. Categories of allergic comorbidities

<table>
<thead>
<tr>
<th>Category</th>
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<tbody>
<tr>
<td>No Disease</td>
</tr>
<tr>
<td>Rhinitis</td>
</tr>
<tr>
<td>Asthma</td>
</tr>
<tr>
<td>Eczema</td>
</tr>
<tr>
<td>Eczema + Asthma</td>
</tr>
<tr>
<td>Eczema + Rhinitis</td>
</tr>
<tr>
<td>Asthma + Rhinitis</td>
</tr>
<tr>
<td>Eczema + Asthma + Rhinitis</td>
</tr>
</tbody>
</table>

To estimate the single and combined effects of the risk factors, FLG variants and
repeated measures of allergic sensitization, on allergic disorders, generalized estimating
equations (GEE) were used when estimating the covariance matrix to account for
correlated observations and the within-child effect of the repeated measurements [88].
Risk ratios (RR) and their 95% confidence intervals (95% CIs) were estimated by
applying log-binomial regression models using the GENMOD procedure in SAS 9.3
(SAS, Gary, NC, USA). The “no allergic disorder” category formed the reference
category. The main effects of FLG variants and allergic sensitization on the risk for
single allergic disorders and comorbidities were estimated separately (single risk factor
models). In a full model, the two main plus their interaction effect (FLG variants ×
allergic sensitization) were estimated. We repeated the aforementioned analysis while
excluding the 1-or-2 years follow-up data from the GEE analysis to estimate whether
possible misclassifications of phenotypes at early life biased our results. In the case of a
possibly statistically significant interaction ($P_{interaction} < 0.1$), the combined effect was
estimated as follows: \( RR = \exp[(\beta_1 \times \text{allergic sensitization}) + (\beta_2 \times FLG \text{ variants}) + (\beta_3 \times \text{allergic sensitization} \times FLG \text{ variants})] \). The term ‘combined effect’ was used to
describe the joint impact of two individual risk factors plus their interaction on the occurrence of the outcome. In all GEE models, sex and age at follow-up were included as potential confounders. To evaluate statistical significance of main effects of covariates (not interaction terms), a $P$-value $\leq 0.05$ was used as indicator of statistical significance.

To determine the proportion of the different allergic morbidities/comorbidities related to $FLG$ variants and allergic sensitization, population attributable fractions (PAFs) were estimated using the following formula: $\text{PAF} = p_c \times [(\text{RR} - 1)/\text{RR}]$, where $p_c$ is the proportion of diseased-individuals that are exposed [89, 90]. This approach yields valid PAF estimates in the presence of confounding and/or effect modification, only when using adjusted RRIs [89, 90]. In case of statistically significant interactions between allergic sensitization and $FLG$ variants, the RRIs for the combined effect were used to estimate PAFs associated with both risk factors. If no interaction was detected, adjusted RRIs were used to estimate separate PAFs for each of the two risk factors and an overall PAF was obtained using the following formula: $\text{PAF}_{\text{overall}} = 1 - [(1 - \text{PAF}_{\text{allergic sensitization}}) \times (1 - \text{PAF}_{FLG \text{ variants}})]$ [89].

2.5.3 Specific Aim 3: Statistical Analysis

**SA3:** To associate DNA methylation across the EDC genomic region with eczema status (H4).

Although genome-wide DNA methylation profiles were available, I restricted the analysis to 256 CpG sites that span the EDC genomic region (location on human chromosome 1: 151958685 – 153628983). After performing quality control measures on DNA methylation data, Beta-values presented as the proportion of methylated (M) over
methylated (M) and unmethylated (U) sites ($\text{Beta-value} = \frac{M}{c+M+U}$, with c being constant to prevent dividing by zero) were calculated for each CpG site [87]. Beta-values provide intuitive biological interpretation (i.e., % methylation); however, (i) being constrained between values of 0 (unmethylated) and 1 (completely methylated) and (ii) demonstrating high heteroscedasticity in the lower and upper ends of the methylation range has raised concerns on the validity of this measure when performing statistical analysis [91]. The conversion of Beta-values to $M$-values ($M$-value = $\log_2(Beta\text{-value}/[1 – Beta\text{-value}])$) has been shown to overcome the limitations of beta-values [91]. Moreover, the use of $M$-values, when performing statistical analysis, demonstrated increased reliability in terms of detecting differentially methylated CpG sites; however, $M$-values cannot be directly interpreted [91, 92]. Therefore, following previous recommendation [91], we used $M$-values when performing the statistical analysis and reported the statistics at the scale of Beta-values.

All statistical analyses were conducted using SAS® version 9.3 (SAS Institute, Cary, NC, USA). In the discovery cohort, linear regression was used to test the association between DNA methylation ($M$-values) of all CpG sites spanning the EDC region with eczema status adjusting for sex and cell-type composition. To minimize false-positive findings identified in 256 models, CpG sites with a false discovery rate (FDR) adjusted p-value < 0.1 were selected and considered as potentially associated with eczema. The selected CpG sites, in the discovery cohort, were taken forward and tested for their possible association with eczema status in replication cohorts I and II. Then, the selected CpG sites were tested for association with eczema in males and females separately.
CHAPTER 3

RESULTS I

MANUSCRIPT # 1: ASSOCIATION OF FILAGGRIN VARIANTS WITH ASTHMA AND RHINITIS: IS ECZEMA OR ALLERGIC SENSITIZATION STATUS AN EFFECT MODIFIER?¹

3.1 Abstract

**Background:** Association of filagrin (*FLG*) variants with asthma and rhinitis have shown to be modulated by eczema status. However, it is unknown whether allergic sensitization status modifies this association. The aim of this study was to determine whether *FLG* variants need eczema and/or allergic sensitization as a necessary component to execute its adverse effect on coexisting and subsequent asthma and rhinitis.

**Methods:** Repeated measurements of asthma, rhinitis, eczema, and allergic sensitization (documented by skin prick tests) at ages 1, 2, 4, 10, and 18 years were ascertained in the Isle of Wight birth cohort (n = 1,456). *FLG* haploinsufficiency was defined as having at least the minor allele of R501X, 2282del4, or S3247X variants. Log binomial regression models were used to test associations and statistical interactions.

**Results:** *FLG* variants increased the risk of asthma (RR = 1.39, 95% CI: 1.06 – 1.80) and rhinitis (RR = 1.37, 95% CI: 1.16 – 1.63). In delayed effect models, ‘*FLG* variants plus allergic sensitization’ and ‘*FLG* variants plus eczema’ increased the risk of subsequent asthma by 4.93-fold (95% CI: 3.61 – 6.71) and 3.33-fold (95% CI: 2.45 – 4.51), respectively, during the first 18 years of life. In contrast, neither eczema nor allergic sensitization in combination with *FLG* variants increased the risk of later rhinitis.

**Conclusions:** Allergic sensitization and eczema modulated the association between *FLG* variants and asthma, but not rhinitis. Results of our study imply that the mechanisms and pathways through which *FLG* variants predispose to increased risk of asthma and rhinitis may be different.
3.2 Introduction

The burden associated with allergic disorders, including eczema, asthma, and rhinitis is substantial and of public health importance. Prior investigations indicated that a complex interplay between genetic, immunological, and environmental factors contributes to the development and maintenance of such allergic manifestations. Until recently, etiologic research focused on the role of immune dysregulation in the pathogenesis of allergic disorders; however, a shift in the research paradigm towards understanding the contribution of a defective epidermal barrier rapidly emerged after the discovery of loss-of-function variants in the filaggrin gene \((FLG)\) [53, 56, 60]. Filaggrin haploinsufficiency, defined as partial or complete loss of filaggrin (filament-aggregating protein) protein, is associated with the development of an impaired epidermal barrier that is characterized by increased allergen penetration and water permeability [93].

Thus far, \(FLG\) variants are the most replicated and strongest genetic risk factor for eczema [59]. Also, \(FLG\) variants are considered to be associated with asthma and rhinitis [66]. Of importance is that the association between \(FLG\) variants and asthma is stronger in the presence of eczema [59, 66]. However, this possible effect modification by eczema was not demonstrated for the association between \(FLG\) variants and rhinitis [66]. Mechanisms that underlie the association of \(FLG\) variants with asthma and rhinitis are not well understood since \(FLG\) is not expressed in the upper or lower airway epithelium [94, 95]. A proposed pathway is that cutaneous sensitization, facilitated by \(FLG\) variants, may lead to local and systematic inflammation at distant organs (i.e., lung and nasal tissues) [56, 60].
While the concept that eczema leads to asthma in children who have a loss-of-function variants in the *FLG* gene is attractive, research in this area has neglected the role of allergic sensitization, defined as the propensity to produce immunoglobulin E (IgE) antibodies responses to environmental and food antigens [96]. Only one study based on the German Multicenter Allergy Study (MAS) birth cohort addressed allergic sensitization and reported an interaction between *FLG* variants and food sensitization in the pathogenesis of asthma among children with eczema [97]. Hence, further investigations on whether the associations of *FLG* variants with asthma and rhinitis are modified by eczema and/or allergic sensitization are needed so that preventive efforts can be directed to either clinical management of eczema or allergic sensitization or both.

We hypothesized that allergic sensitization, rather than eczema status, modifies the association of *FLG* variants with asthma and rhinitis. Therefore, statistical interactions (*FLG* variants × allergic sensitization and *FLG* variants × eczema) were tested to determine whether *FLG* variants plus allergic sensitization and/or *FLG* variants plus eczema jointly increased the risk of asthma and/or rhinitis. To this end, analyzing longitudinal data from the Isle of Wight birth cohort covering childhood and adolescence prospectively enabled us to determine whether *FLG* variants need allergic sensitization or eczema as a necessary component to execute its adverse effect on coexisting and subsequent asthma and rhinitis.
3.3 METHODS

3.3.1 STUDY DESIGN AND PARTICIPANTS

An unselected whole population birth cohort (n = 1,536) was recruited in 1989 in the Isle of Wight, UK, to prospectively study the natural history of allergic conditions. After exclusion of adoptions, perinatal deaths, and refusal for follow-up, 1,456 (95%) children were enrolled, with follow-up assessments conducted at 1, 2, 4, 10, and 18 years of age. Ethics approvals were obtained from the Isle of Wight Local Research Ethics Committee (NRES Committee South Central – Southampton B) at recruitment and for the subsequent follow-ups (06/Q1701/34). At each follow-up, validated questionnaires, including the International Study of Asthma and Allergy in Childhood (ISAAC) questionnaire [4], were completed on allergic disorders plus demographic attributes and exposures to environmental factors.

3.3.2 PHENOTYPES

In all assessments of the Isle of Wight birth cohort, eczema was defined as chronic or chronically relapsing, itchy dermatitis lasting more than 6 weeks with characteristic morphology and distribution [83], following Hanifin and Rajka criteria [84]. For asthma, at the 1, 2, and 4-year follow-ups, the medical investigator determined the presence of asthma based on wheeze frequency over the last 12 months and treatment given for asthma or asthma related symptoms. At the 10 and 18 year follow-ups, asthma was defined as having “ever had asthma” and either “wheezing or whistling in the chest in the last 12 months” or “current treatment for asthma”, using ISAAC questionnaire [4]. Rhinitis was defined by a positive response to: ‘In the past 12 months have you had a
problem with sneezing, or a runny or a blocked nose when you did not have a cold or the flu?’ [33] Since the 1-year and 2-year follow-up data on eczema, asthma, and rhinitis were collected in a relatively small time window, we combined them for analytic purposes (reported as 1-or-2 years).

To determine allergic sensitization status, skin prick testing (SPT) at ages 1 and 2 years was performed on children with any symptoms of eczema, asthma, or rhinitis. We combined SPT results for ages 1 and 2 years since they occurred within a short time period and will henceforth refer to this as SPT at 1-or-2 years. At 4, 10, and 18 years, regardless of symptoms, SPT was performed on most children attending the research center to a standard battery of common allergens (ALK-Albello, Horsholm, Denmark). Inhalant allergens tested were house dust mite, cat, dog, *Alternaria alternata*, *Cladosporium herbarium*, grass pollen mix, and tree pollen mix. Food allergens tested were cows’ milk, soya, hens’ egg, peanut and cod. Positive and negative controls were included. Allergic sensitisation was defined by having a positive SPT to at least one allergen test with mean wheal diameter of 3 mm greater than the negative control. Since allergic sensitization is a dynamic rather than a completely stable phenotype, we used the concurrent and/or preceding status and thus allowed the risk to change over time.

3.3.3 FLG Genotyping

Blood and/or saliva samples were collected at ages 10 and/or 18 years from which genomic DNA was isolated. DNA samples were interrogated using GoldenGate Genotyping Assays (Illumina, Inc, SanDiego, CA) on the BeadXpressVeracode platform (Illumina, Inc, SanDiego, CA) per Illumina’s protocol. Individuals carrying the minor
allele for at least one of the FLG variants R501X, 2282del4, or S3247X were classified as having filaggrin haploinsufficiency. The R2447X variant was also genotyped, but none of the study participants carried the minor allele. Detailed information on genotyping is provided by Ziyab et al. [79].

3.3.4 Statistical Analysis

Since the proposed effect modifiers (i.e., allergic sensitization and eczema) and the two outcomes (i.e., asthma and rhinitis) were measured repeatedly at ages 1-or-2, 4, 10, and 18 years, we applied the generalized estimating equation (GEE) method with first-order autoregressive covariance matrix to account for the correlated observations and the within-child effect of the repeated measurements [88]. Risk ratios (RR) and their 95% confidence intervals (95% CIs) were estimated by applying log-binomial regression models using the GENMOD procedure in SAS 9.3 (SAS, Gary, NC, USA). To determine whether the RR of the association of FLG variants with asthma and rhinitis differ according to the presence or absences of the proposed effect modifiers, we evaluated the association in the total study sample and sub-samples based on the status of the possible effect modifiers.

Interaction terms, on a multiplicative scale, were used to test the additional effect of two co-occurring risk factors on the health outcomes above and beyond their individual effects. Also, to evaluate whether the effect of FLG variants across levels of the effect modifiers were statistically significantly different, we included interaction terms in separate regression models (model 1: FLG variants × eczema and model 2: FLG variants × allergic sensitization). To decipher which interactive effect was more
pronounced, a regression model including both interaction terms (model 3: FLG variants × eczema and FLG variants × allergic sensitization) was evaluated. Henceforth, we refer to models 1, 2, and 3 as the ‘concurrent models’ since the effect modifiers and the outcomes coexisted at the same time. In the case of a possibly statistically significant interaction term ($P_{\text{interaction}} < 0.1$), the “combined effect”, referring to the joint impact of two individual risk factors plus their interaction on the occurrence of the outcome, was estimated. We repeated the previous analyses while stratifying by sex to determine if sex-specific effects existed. In all GEE models, sex and age at follow-up were included as potential confounders.

In addition to the concurrent models, we tested the interactions in ‘delayed effect’ models to investigate whether the time-order of risk and response supports the concurrent model findings. The delayed effect analysis helps to determine whether the interaction effects contribute to the developmental process of asthma and rhinitis. To this end, we analyzed whether FLG variants interact with preceding allergic sensitization or eczema on the risk of subsequent asthma and rhinitis. For example, the interaction of FLG variants with allergic sensitization status at age 1-or-2 years on the risk of asthma at age 4 years (FLG variants × SPT results at 1-or-2 years $\rightarrow$ asthma at 4 years) was evaluated. The prevalence of the clinical phenotype (e.g., asthma at 4), which was used in the delayed effect models, included new occurrences and persistence of the disease. Therefore, we constructed three delayed periods (1-or-2 to 4 years, 4 to 10 years, and 10 to 18 years) to determine if the presence of both FLG variants and preceding allergic sensitization or preceding eczema influences the positive transition, defined as the change
in disease status from disease-free to diseased, and the persistence of the disease in two consecutive follow-ups [37].

3.4 Results

3.4.1 Description of Study Population

Of the 1,456 children enrolled in the study, 1,377 were followed-up at age 1-or-2 years, 1,214 at 4 years, 1,368 at 10 years, and 1,309 at 18 years. The period prevalence ranged between 11.9% to 14.2% for eczema, 14.3% to 17.7% for asthma, and 5.6% to 35.8% for rhinitis (table 3.1). At age 1-or-2 years, SPTs were performed on symptomatic children (n = 515), of which 20.6% had at least one positive SPT response. At all other ages, the majority of participants underwent SPTs with the proportion of positive SPTs being 19.4% at 4 years, 26.9% at 10 years, and 41.4% at 18 years of age. Genotype frequencies of FLG variants (table 3.1) were concordant with Hardy-Weinberg equilibrium.

3.4.2 FLG Variants and Asthma

In the total study sample, FLG variants increased the risk of repeated occurrence of asthma from 1-or-2 to 18 years of age (RR = 1.39, 95% CI: 1.06 – 1.80; figure 3.1). In the presence of eczema, the effect size increased (RR = 1.56, 95% CI: 1.10 – 2.21); in the absence of eczema, the magnitude of the association between FLG variants and asthma was reduced and lost statistical significance (RR = 1.26, 95% CI: 0.93 – 1.69). This association was noticeably modified when stratifying based on the presence (RR = 1.49,
95% CI: 1.14 – 1.94) or absence (RR = 0.76, 95% CI: 0.48 – 1.21) of allergic sensitization.

To determine whether the observed differences in the associations were statistically significant, statistical interactions on a multiplicative scale were evaluated in concurrent models. There was no evidence for an interaction between FLG variants and eczema on the risk for asthma (interaction effect RR = 1.18, 95% CI: 0.78 – 1.79, \( P_{\text{interaction}} = 0.429 \); figure 3.1). However, a statistically significant interaction was found between FLG variants and allergic sensitization on the risk for asthma (interaction effect RR = 2.00, 95% CI: 1.22 – 3.28, \( P_{\text{interaction}} = 0.006 \); figure 3.1). The combined effect of FLG variants and allergic sensitization increased the risk of asthma by 3.63-fold (95% CI: 2.81 – 4.70). Furthermore, to determine which interactive effect is more important, we simultaneously included both interaction terms in one regression model. Results showed that the interaction between FLG variants and allergic sensitization remained statistically significant after adjusting for the interactive effect between FLG variants and eczema (data not shown). However, the latter did not gain statistical significance.

We retested the results of the aforementioned concurrent models, using delayed effect models. In these models we used the preceding cumulative SPTs results (or eczema status), i.e. all SPTs results (or eczema status) from assessments that are prior to the outcome assessment, when testing the interaction between FLG variants and allergic sensitization (or eczema) on the risk of subsequent asthma. The results of the delayed effect models show that the combined effect of preceding allergic sensitization with FLG variants increased the risk of asthma in the next exam at each delay period (figure 3.2a). For instance, the presence of both FLG variants plus positive SPT at 1-or-2 and/or 4 years
increased the risk of asthma at 10 years (RR = 7.22, 95% CI: 4.84 – 10.80). Although results of the concurrent model (figure 3.1) did not show an interaction between FLG variants and eczema, the delayed effect models demonstrated interactive effects between FLG variants and preceding eczema on the risk of subsequent asthma (figure 3.2b). The repeated measurement analysis shows that the combined effect of FLG variants and preceding eczema, on average, increased the risk of later asthma during the first 18 years of life (RR = 3.33, 95% CI: 2.45 – 4.51).

To further understand whether the observed interactions between FLG variants and preceding allergic sensitization and eczema influence new occurrence and/or persistence of asthma, we calculated the proportions of positive transitions (defined as the change in disease status from disease-free to diseased in two consecutive follow-ups) and persistence among those with and without both risk factors (table 3.2). The proportion of positive transition was more frequent among those with FLG variants and allergic sensitization as compared to those without both risk factors across all transition periods. For instance, in the presence of both FLG variants and allergic sensitization, 5/16 (31.3%) of those without asthma at age 4 years developed asthma at age 10 years; whereas, only 23/532 (4.3%) of those without both risk factors developed asthma at 10 years. Furthermore, the presence of both risk factors influenced the persistence of asthma (table 3.2). For example, 18/21 (85.7%) of participants with both risk factors who had asthma at ages 10 years continued to have asthma at 18 years, as compared to 25/44 (56.8%) among those without the two risk factors. Similarly, the presence of both FLG variants and preceding eczema influenced the positive transition and persistence of asthma.
3.4.3 FLG Variants and Rhinitis

FLG variants were associated with increased risk for rhinitis from age 1-or-2 to 18 years in the total study sample of repeated measurements (RR = 1.37, 95% CI: 1.16 – 1.63; figure 3.3). In both the presence (RR = 1.57, 95% CI: 1.23 – 2.00) and absence (RR = 1.28, 95% CI: 1.04 – 1.57) of eczema, the association between FLG variants and rhinitis was statistically significant. However, we observed a statistically significant association between FLG variants and rhinitis only in the presence of allergic sensitization (RR = 1.34, 95% CI: 1.17 – 1.53), and not in its absence (RR = 1.13, 95% CI: 0.82 – 1.55; figure 3.3).

To test if the observed heterogeneous effects across the levels of the potential effect modifiers are statistically different, we tested multiplicative statistical interactions in concurrent models. There was no evidence for an interactive effect neither between FLG variants and eczema (interaction effect RR = 1.15, 95% CI: 0.83 – 1.60, \( P_{\text{interaction}} = 0.409 \); figure 3.3) nor between FLG variants and allergic sensitization (interaction effect RR = 1.07, 95% CI: 0.76 – 1.51, \( P_{\text{interaction}} = 0.698 \); figure 3.3) on the risk for rhinitis. Concordant with results of concurrent models, the delayed effect models did not show any interactions between FLG variants and preceding allergic sensitization or eczema on the risk of subsequent rhinitis (figure 3.4).

3.4.4 Sex Differences and Time Trends

Separate analyses for each sex were performed and the results for boys and girls were similar (data not shown) and in agreement with the results of analyzing both sexes together in the repeated measurement (GEE) analysis. Furthermore, at each follow-up we
analyzed boys and girls separately to determine whether the effects differ across both time and sex. The obtained results indicated that neither age of exam nor sex influenced our results.

3.5 DISCUSSION

The objective of this study was to determine whether eczema and/or allergic sensitization status act as effect modifiers for the association of FLG variants with asthma and rhinitis. In this study, FLG variants increased the risk of asthma and rhinitis during the first 18 years of life in the total study sample. In concurrent effect models, interaction between FLG variants and allergic sensitization resulted in a combined effect that increased the risk of coexisting asthma by 3.636-fold. Delayed effect models, which take the time order of risk factors and disease occurrences into account, supported the findings of the concurrent effect models regarding allergic sensitization and FLG variants. In addition, the delayed effect models showed that FLG variants interact with preceding eczema status on the development of subsequent asthma, indicating that both allergic sensitization and eczema act as effect modifiers for the association between FLG variants and asthma. In contrast, neither eczema nor allergic sensitization modified the association of FLG variants with rhinitis in a statistically significant manner.

The association between FLG variants and asthma among those with eczema was first reported by Palmer et al. [53]. Subsequently, several studies have replicated this association and added a possible association between FLG variants and rhinitis [59, 66]. Results of our study further support the general agreement that FLG variants are associated with asthma in the presence of eczema. We further demonstrated that both
preceding allergic sensitization and eczema modified the association between \textit{FLG} variant and asthma through multiplicative interaction (figure 3.2). These findings indicate that \textit{FLG} variants need eczema or allergic sensitization to execute their adverse effects on asthma. The finding of an interaction between \textit{FLG} variants and allergic sensitization in this study improves our understanding of the possible link between genetics of the epidermal barrier (i.e., \textit{FLG} variants) and a respiratory disorder (i.e., asthma). Such an observation further supports the hypothesis that cutaneous sensitization priming, facilitated by \textit{FLG} variants, may migrate to the airways and cause local and systematic inflammation [98]. Although prior studies have widely suggested such a pathway, the majority of previous investigations did not take the status of allergic sensitization into account. An exception is the study by Marenholz \textit{et al.}, which showed an interaction between \textit{FLG} variants and food sensitization on the risk of asthma in children with eczema [97], however, their analytical sample was restricted to children with eczema.

In the delayed effect models we used the prevalence of asthma as the outcome, which does not distinguish between new occurrences and persistence of asthma. Therefore, in additional descriptive analysis, we demonstrated that the presence of both \textit{FLG} variants and preceding allergic sensitization or preceding eczema influenced the new occurrences (positive transition) and persistence of asthma. Hence, our data suggest that the presence of both risk factors (‘\textit{FLG} variants and preceding allergic sensitization’ or ‘\textit{FLG} variants and preceding eczema’) plays an important role in the development and persistence of asthma.

Attempts using the composite of ‘atopic asthma’ phenotype were conducted to investigate whether allergic sensitization is the link between \textit{FLG} variants and asthma.
Inconsistent results for the association of FLG variants with ‘atopic asthma’ have been reported [62, 64, 99, 100]. These inconsistencies could be attributed to the use of the composite ‘atopic asthma’ phenotype, which does not clearly define the reference (no risk) group. By taking this approach investigators tend to group ‘non-atopic asthmatics’ and ‘atopics without asthma’ participants together in the reference group. Therefore, when assessing the association of FLG variants with ‘atopic asthma’ one should be aware that the reference group could include atopic participants, which could result in distorting assessments. The advantage of our approach, using two separate risk factors, over the composite ‘atopic asthma’ phenotyping, is that we were able to estimate the additional effect due to interaction (FLG variants × allergic sensitization) that is above and beyond their independent effects. Hence, allowing us to determine whether the combined effect of both risk factors is more than just the multiplication of their independent effects.

There is conflicting evidence in the literature as to whether the association between FLG variants and rhinitis is modulated by eczema status. The majority of previous studies found a stronger association between FLG variants and rhinitis in the presence of coexisting eczema when compared to their association in the absence of concurrent eczema [62, 63, 65, 66, 101]. Another investigation reported an association between FLG variants and ‘persistent allergic rhinitis’ phenotype [99]. Controversially, however, a significant association has also been reported between FLG variants and the composite ‘allergic rhinitis’ phenotype adjusted for eczema status [64]. Our analyses did not reveal interaction between eczema ($P_{interaction} = 0.409$) or allergic sensitization ($P_{interaction} = 0.698$) with FLG variants on the risk of rhinitis in the repeated measurement analysis. Our results of an interactive effect between FLG variants and allergic
sensitization on the risk of asthma and the lack of such interaction on the risk of rhinitis suggest that the pathway through which FLG variants predispose to asthma and rhinitis may be different. For instance, the expression of filaggrin protein was not detected in the human bronchial epithelium; however, it has been shown that filaggrin is expressed in the nasal vestibule [64, 94]. That said, the extent to which filaggrin expression in the nasal vestibular lining influence the development of rhinitis is yet to be investigated by future studies.

Major strengths of our prospective, 18-year study are the repeated phenotyping, objective assessments of allergic sensitization status, and the low loss to follow-up (ranged from 5% to 17%). Moreover, the majority (80%) of the study participants were genotyped for FLG variants common among populations of European ancestry [57]. We showed, previously, that the genotyped study participants did not differ from the total cohort with regard to multiple characteristics [79]. Hence, there is no indication of selection bias that could pose a threat to the validity of our study. Misclassification of eczema cases is minimal since a high proportion of subjects showed typical manifestation of eczema in the usual locations (antecubital or popliteal fossae, ankles, face or neck for 97% at 1 year, 91% at 2 years, 75% at 4 years, 86% at 10 years and 76% at 18 years) [25].

Potential limitations are the definition of asthma and rhinitis symptoms in early life. Our asthma conclusion at ages 10 and 18 years followed the ISAAC criteria [4], which was at that time not available for assessments at age 1, 2, and 4 years. Although slightly different methods were used to define asthma, in a previous report we have shown that the minor change in asthma definition over time did not influence the validity
of our asthma classification [37]. In addition, since it is difficult to differentiate between infectious and other forms of rhinitis in infancy, the elevated prevalence at 1-or-2 years might have been influenced by misclassifying ‘viral induced infectious rhinitis’ [33]. However, by applying repeated measurements analysis including assessments later in childhood and adolescence, we believe that the influence of the possible misclassification of rhinitis at 1-or-2 years on the overall results of the study is minimal, since similar results were obtained when we reran our analyses excluding the 1-or-2 year data. Moreover, since SPTs at age 1-or-2 years were performed on symptomatic children, we speculate that selection bias leading to underestimated RRs might have been induced. Although proportions of SPT positivity at ages 1-or-2 years (20.6%) and 4 years (19.6%; SPT performed irrespective of symptoms) were similar, RRs associating allergic sensitization at age 1-or-2 years with outcomes (i.e., eczema, asthma, and rhinitis) at the same age tended to be smaller than RRs relating allergic sensitization at age 4 years with outcomes at the same age (data not shown), since also the reference group had some symptoms. Hence, the estimated RRs at age 1-or-2 might be underestimated; however, the extent of the possible selection bias is minimal since our results did not noticeably change when we excluded the 1-or-2 years follow-up data from the analyses. Such observations further demonstrate the robustness of the GEE method in providing population averaged estimates (RRs) while accounting for the within-subject correlations of repeated measurements. The use of a slightly relaxed statistical significance threshold ($P_{\text{Interaction}} < 0.1$) for detecting interaction-effects is further supported by the fact that majority of epidemiologic studies lack sufficient power to detect higher order-terms [102].
In conclusion, our results indicate that allergic sensitization and eczema status are effect modifiers of the association between FLG variants and asthma. The combined effects of ‘preceding allergic sensitization and FLG variants’ and ‘preceding eczema and FLG variants’ increased the risk of subsequent asthma by 4.93-fold and 3.33-fold, respectively, during the first 18 years of life. These findings suggest that FLG variants need allergic sensitization or eczema to execute its adverse effects on asthma. In contrast, neither eczema nor allergic sensitization status statistically significantly modulated the association between FLG variants and rhinitis. Hence, results of our study suggest that the mechanisms and pathways through which FLG variants, representing impaired epidermal barrier, predispose to increased risk of asthma and rhinitis may be different. Future studies confirming our reported observations and exploring the differential etiological pathways underlying the development of asthma and rhinitis are needed to better identify and stratify those who share similar risk characteristics.

3.6 Acknowledgements

We thank participants and their families who have helped us with this project over the last two decades. We would like to acknowledge the help of all the staff at The David Hide Asthma and Allergy Research Centre in undertaking the 18 year and previous assessments of 1989 Isle of Wight birth cohort. The 18 year assessment of the IOW birth cohort and future analyses were funded by grants from the National Institute of Health, USA (R01 HL082925 and R01AI091905).
Figure 3.1. Concurrent association of \textit{FLG} variants with asthma in total study sample and sub-samples: longitudinal analysis covering 1- or 2- to 18 years of age. The prevalence of asthma among those with filaggrin loss-of-function (\textit{FLG LOF}) variants and those with wild-type genotype (\textit{FLG WT}) is shown. RR: Risk ratio; 95\% CI: 95\% confidence interval; \(k\): number of repeated measurements.
Figure. 3.2. a) Delayed effect models exploring the individual and combined effects of FLG variants and preceding allergic sensitization status on asthma development. Independent effects of FLG variants (SPT - & FLG LOF), positive SPT (SPT + & FLG WT), and their combined effect (SPT + & FLG LOF) in the development of asthma are shown for different ages. We modeled the effect of preceding SPT results on later asthma. b) Delayed effect models exploring the individual and combined effects of FLG variants and preceding eczema status on asthma development. Independent effects of FLG variants (Eczema - & FLG LOF), eczema (Eczema + & FLG WT), and their combined effect (Eczema + & FLG LOF) in the development of asthma are shown for different ages. We modeled the effect of preceding eczema results on later asthma. SPT +: positive SPT, SPT -: negative SPT; Eczema +: positive eczema diagnosis, Eczema -: negative eczema diagnosis; FLG LOF: filaggrin loss-of-function; FLG WT: filaggrin wild-type genotype; n = number of children; k= number of repeated measurements. The asterisk (*) and the double dagger (‡) indicate that the interaction effect (a: SPT × FLG; b: Eczema × FLG) is possibly statistically significantly different from the null value of 1 (*: PInteraction < 0.1; ‡: PInteraction < 0.05); suggesting interaction on multiplicative scale. The dagger (†) indicates that the risk estimate (RR) of the given column is statistically significantly (P < 0.05) different from the column without both risk factors (a: SPT - & FLG WT; b: Eczema & FLG WT). The presence of both an asterisk (*) or double dagger (‡) and dagger (†) signifies that the combined effect of the two risk factors is above and beyond their individual effects.
Figure 3.3. Concurrent association of FLG variants with rhinitis in total study sample and sub-samples: longitudinal analysis covering 1-or-2 to 18 years of age. The prevalence of rhinitis among those with filaggrin loss-of-function (FLG LOF) variants and those with wild-type genotype (FLG WT) is shown. RR: Risk ratio; 95% CI: 95% confidence interval; k= number of repeated measurements.
Figure 3.4. **a)** Delayed effect models exploring the individual and combined effects of \textit{FLG} variants and preceding allergic sensitization status on rhinitis development. Independent effects of \textit{FLG} variants (SPT - & \textit{FLG} LOF), positive SPT (SPT + & \textit{FLG} WT), and their combined effect (SPT + & \textit{FLG} LOF) in the development of rhinitis are shown for different ages. We modeled the effect of preceding SPT results on later rhinitis. **b)** Delayed effect models exploring the individual and combined effects of \textit{FLG} variants and preceding eczema status on rhinitis development. Independent effects of \textit{FLG} variants (Eczema - & \textit{FLG} LOF), eczema (Eczema + & \textit{FLG} WT), and their combined effect (Eczema + & \textit{FLG} LOF) in the development of rhinitis are shown for different ages. We modeled the effect of preceding eczema results on later rhinitis. SPT +: positive SPT, SPT -: negative SPT; Eczema +: positive eczema diagnosis, Eczema -: negative eczema diagnosis; \textit{FLG} LOF: filaggrin loss-of-function; \textit{FLG} WT: filaggrin wild-type genotype; n = number of children; k= number of repeated measurements. The dagger (†) indicates that the risk estimate (RR) of the given column is statistically significantly ($P < 0.05$) different from the column without both risk factors (**a**: SPT - & \textit{FLG} WT; **b**: Eczema & \textit{FLG} WT).
Table 3.1. Characteristics of study population

<table>
<thead>
<tr>
<th>Attributes</th>
<th>% (n/total)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sex</strong></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>51.2 (786/1536)</td>
</tr>
<tr>
<td>Eczema at 1-or-2 years</td>
<td>14.2 (196/1377)</td>
</tr>
<tr>
<td>4 years</td>
<td>11.9 (145/1214)</td>
</tr>
<tr>
<td>10 years</td>
<td>13.7 (186/1359)</td>
</tr>
<tr>
<td>18 years</td>
<td>12.3 (161/1307)</td>
</tr>
<tr>
<td>Asthma at 1-or-2 years</td>
<td>14.3 (197/1377)</td>
</tr>
<tr>
<td>4 years</td>
<td>14.9 (181/1214)</td>
</tr>
<tr>
<td>10 years</td>
<td>14.7 (201/1368)</td>
</tr>
<tr>
<td>18 years</td>
<td>17.7 (231/1305)</td>
</tr>
<tr>
<td>Rhinitis at 1-or-2 years</td>
<td>15.8 (217/1377)</td>
</tr>
<tr>
<td>4 years</td>
<td>5.6 (65/1214)</td>
</tr>
<tr>
<td>10 years</td>
<td>22.6 (308/1362)</td>
</tr>
<tr>
<td>18 years</td>
<td>35.8 (468/1309)</td>
</tr>
<tr>
<td>FLG variants*</td>
<td></td>
</tr>
<tr>
<td>R501X</td>
<td>4.1 (47/1161)</td>
</tr>
<tr>
<td>2282del4</td>
<td>4.6 (54/1168)</td>
</tr>
<tr>
<td>S3247X</td>
<td>1.6 (18/1165)</td>
</tr>
<tr>
<td>Combined</td>
<td>10.3 (118/1150)</td>
</tr>
<tr>
<td>Allergic sensitization† at</td>
<td></td>
</tr>
<tr>
<td>1-or-2 years</td>
<td>20.6 (106/515)</td>
</tr>
<tr>
<td>4 years</td>
<td>19.6 (192/982)</td>
</tr>
<tr>
<td>10 years</td>
<td>26.9 (279/1036)</td>
</tr>
<tr>
<td>18 years</td>
<td>41.4 (353/853)</td>
</tr>
</tbody>
</table>

*Analyses were conducted using the combined carrier frequency of 10.3%.
† Presence of allergic sensitization was defined by having on or more positive skin prick test result at the respective assessment.
Table 3.2. Prevalence, positive transition, and persistence of asthma stratified based on FLG variants and preceding allergic sensitization and eczema status: comparing proportions of those with both risk factors to those without the risk factors

<table>
<thead>
<tr>
<th></th>
<th>Asthma % (n/total)</th>
<th>Prevalence</th>
<th>Positive transition</th>
<th>Persistence</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Age 4 years</td>
<td>Age 10 years</td>
<td>Age 18 years</td>
<td></td>
</tr>
<tr>
<td>FLG WT &amp; SPT –</td>
<td>18.4 (57/310)</td>
<td>7.9 (47/595)</td>
<td>10.3 (62/604)</td>
<td></td>
</tr>
<tr>
<td>FLG LOF &amp; SPT +</td>
<td>50.0 (9/18)</td>
<td>60.0 (18/30)</td>
<td>54.8 (23/42)</td>
<td></td>
</tr>
<tr>
<td>p-value*</td>
<td>0.003</td>
<td>&lt; 0.001</td>
<td>&lt; 0.001</td>
<td></td>
</tr>
<tr>
<td>FLG WT &amp; Eczema –</td>
<td>11.9 (86/725)</td>
<td>11.2 (91/811)</td>
<td>16.0 (118/739)</td>
<td></td>
</tr>
<tr>
<td>FLG LOF &amp; Eczema +</td>
<td>52.0 (5/6)</td>
<td>51.5 (17/33)</td>
<td>51.3 (20/39)</td>
<td></td>
</tr>
<tr>
<td>p-value*</td>
<td>&lt; 0.001</td>
<td>&lt; 0.001</td>
<td>&lt; 0.001</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>1-or-2 to 4 years</th>
<th>4 to 10 years</th>
<th>10 to 18 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>FLG WT &amp; SPT –</td>
<td>8.5 (19/223)</td>
<td>4.3 (23/532)</td>
<td>6.7 (37/550)</td>
</tr>
<tr>
<td>FLG LOF &amp; SPT +</td>
<td>50.0 (6/12)</td>
<td>31.3 (5/16)</td>
<td>23.8 (5/21)</td>
</tr>
<tr>
<td>p-value*</td>
<td>&lt; 0.001</td>
<td>&lt; 0.001</td>
<td>0.014</td>
</tr>
<tr>
<td>FLG WT &amp; Eczema –</td>
<td>8.5 (53/635)</td>
<td>7.0 (44/632)</td>
<td>9.3 (59/639)</td>
</tr>
<tr>
<td>FLG LOF &amp; Eczema +</td>
<td>42.1 (8/19)</td>
<td>29.4 (5/17)</td>
<td>14.3 (3/21)</td>
</tr>
<tr>
<td>p-value*</td>
<td>&lt; 0.001</td>
<td>0.006</td>
<td>0.437</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>1-or-2 to 4 years</th>
<th>4 to 10 years</th>
<th>10 to 18 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>FLG WT &amp; SPT –</td>
<td>43.7 (38/87)</td>
<td>38.7 (24/62)</td>
<td>56.8 (25/44)</td>
</tr>
<tr>
<td>FLG LOF &amp; SPT +</td>
<td>50.0 (3/6)</td>
<td>92.9 (13/14)</td>
<td>85.7 (18/21)</td>
</tr>
<tr>
<td>p-value*</td>
<td>0.542</td>
<td>&lt; 0.001</td>
<td>0.026</td>
</tr>
<tr>
<td>FLG WT &amp; Eczema –</td>
<td>36.7 (33/90)</td>
<td>44.7 (38/85)</td>
<td>68.3 (56/82)</td>
</tr>
<tr>
<td>FLG LOF &amp; Eczema +</td>
<td>83.3 (5/6)</td>
<td>80.0 (12/15)</td>
<td>94.4 (17/18)</td>
</tr>
<tr>
<td>p-value*</td>
<td>0.031</td>
<td>0.023</td>
<td>0.037</td>
</tr>
</tbody>
</table>

FLG LOF: filaggrin loss-of-function; FLG WT: filaggrin wild-type genotype; SPT -: negative skin prick test result; SPT +: positive skin prick test result.

* p-values comparing the proportions across the two exposure groups were derived from fisher’s exact test.
† Positive transition refers to the change in disease status from asthma-free to asthma in two consecutive assessments.
‡ Persistence refers to the proportion of individuals who had asthma at two consecutive assessments.
CHAPTER 4

RESULTS II

MANUSCRIPT #2: ALLERGIC SENSITIZATION AND FILAGGRIN VARIANTS PREDISPOSE TO THE COMORBIDITY OF ECZEMA, ASTHMA, AND RHINITIS: RESULTS FROM THE ISLE OF WHITE BIRTH COHORT²

4.1 Abstract

**Background:** Allergic sensitization and filaggrin gene (FLG) variants are important risk factors for allergic disorders; however, knowledge on their individual and interactive effects on the coexistence of eczema, asthma, and rhinitis is lacking.

**Objective:** This study aimed at investigating the single and combined effects of allergic sensitization and FLG variants on the development of single and multiple allergic disorders.

**Methods:** The Isle of Wight Birth Cohort (n = 1,456) has been examined at 1, 2, 4, 10, and 18 years of age. Repeated measurements of eczema, asthma, rhinitis, and skin prick tests were available for all follow-ups. FLG variants were genotyped in 1,150 participants. Associations of allergic sensitization and FLG variants with single and multiple allergic disorders were tested in log-binomial regression analysis.

**Results:** The prevalence of eczema-, asthma-, and rhinitis-only ranged from 5.6% to 8.5%, 4.9% to 10.2%, and 2.5% to 20.4%, respectively, during the first 18 years of life. The coexistence of allergic disorders is common, with approximately 2% of the population reporting the comorbidity of “eczema, asthma, and rhinitis” during the study period. In repeated measurement analyses, allergic sensitization and FLG variants, when analyzed separately, were associated with having single and multiple allergic disorders. Of particular significance, their combined effect increased the risk of “eczema and asthma” (RR = 13.67, 95% CI: 7.35 – 25.42), “asthma and rhinitis” (RR = 7.46, 95% CI: 5.07 – 10.98), and “eczema, asthma, and rhinitis” (RR = 23.44, 95% CI: 12.27 – 44.78).

**Conclusions and Clinical Relevance:** The coexistence of allergic disorders is frequent and allergic sensitization and FLG variants jointly increased risk of allergic
comorbidities, which may represent more severe and complex clinical phenotypes. The interactive effect and the elevated proportion of allergic comorbidities associated with allergic sensitization and \textit{FLG} variants emphasize their joint importance in the pathogenesis of allergic disorders.

4.2 \textbf{INTRODUCTION}

Allergic disorders, including eczema, asthma, and rhinitis, pose social and economic burden on individuals, families, and societies [1, 2]. Worldwide, the lifetime prevalence of eczema symptoms (e.g., itchy rash) is estimated to be 15\% to 30\% among children and 2\% to 10\% among adults [19]. Similarly, the prevalence of asthma (up to 25\%) and rhinitis (up to 30\%) is high [16, 29, 32]. Natural history investigations have demonstrated close and complex relationships between these allergic disorders [25, 33, 48, 103-105]. However, mechanisms and pathways underlying their development is an area of ongoing scientific dispute.

The “allergic march” concept suggests that allergic disorders develop in a sequential pattern, starting with eczema in early infancy and later in childhood developing asthma and rhinitis [106]. However, supporting replicable evidence and consensus is far from complete [42, 107, 108]. An opposing concept is the “coexistence” of allergic morbidities [42, 45, 109]. A report based on the German Multicenter Allergy Study showed that single occurrence of eczema early in life does not result in an allergic march, but the coexistence of eczema and wheezing predicted asthma [45]. Results from the BAMSE birth cohort showed that the comorbidity of allergic disorders is frequent during the first 12 years of life [109]. Such observations speak in favor of coexistence of...
allergic morbidities instead of a progressive development. The interrelationship between these conditions and the risk factors that predispose individuals to develop multiple allergic disorders is an open field for research.

Interplay between genetic, environmental, and immunological factors is considered to contribute to the pathogenesis of allergic disorders [110]. Allergic sensitization, defined as the susceptibility to produce immunoglobulin E (IgE) antibodies in response to antigens, is widely-considered as a common thread linking various manifestations of allergic disorders [49, 96]. Likewise, loss-of-function variants in the filaggrin gene (FLG) resulting in an impaired epidermal barrier have rapidly emerged as common risk factor for the development of allergic disorders [60]. However, there is a gap in understanding of the respective contribution of FLG variants and allergic sensitization on the development of allergic comorbidities. Moreover, the joint role of FLG variants and allergic sensitization on the coexistence of eczema, asthma, and rhinitis has not been previously investigated. Using data from the Isle of Wight (IOW) birth cohort, we aimed at determining the single and combined effects of FLG variants and allergic sensitization on the development of single and multiple (coexisting) allergic disorders.

4.3 MATERIALS AND METHODS

4.3.1 STUDY DESIGN AND PARTICIPANTS

An unselected whole population birth cohort (n = 1,536) was recruited in 1989 on the Isle of Wight, UK, to prospectively study the natural history of allergic conditions. After exclusion of adoptions, perinatal deaths, and refusal for follow-up, written informed
consent was obtained from parents to enroll 1,456 (95%) newborns, with follow-up assessments conducted at 1, 2, 4, 10, and 18 years of age. Ethics approvals were obtained from the Isle of Wight Local Research Ethics Committee (now named the National Research Ethics Service, NRES Committee South Central – Southampton B) at recruitment and for the subsequent follow-ups (06/Q1701/34).

4.3.2 Phenotypes

In all assessments of the IOW birth cohort, eczema was defined as chronic or chronically relapsing, itchy dermatitis lasting more than 6 weeks with characteristic morphology and distribution [83], following Hanifin and Rajka criteria [84]. For asthma, at the 1, 2, and 4-year follow-ups, the medical investigator determined the presence of asthma based on wheeze over the last 12 months and treatment given for asthma or asthma related symptoms. At the 10 and 18 year follow-ups, asthma was defined as having “ever had asthma” and either “wheezing or whistling in the chest in the last 12 months” or “current treatment for asthma”, following the International Study of Asthma and Allergies in Childhood (ISAAC) criteria [4]. Rhinitis was defined by a problem with sneezing, or a runny or a blocked nose without a cold or the flu in the last 12 months [33]. Since the 1-year and 2-year follow-up data were collected in a relatively small time window, we combined them for analytic purposes (reported as 1-or-2 years).

To determine allergic sensitization, skin prick tests (SPTs) at ages 1 and 2 years were performed on children with any symptoms of eczema, asthma, or rhinitis. Parallel to the disorders, we combined the SPTs at ages 1 and 2 years (reported as 1-or-2 years). At 4, 10, and 18 years, regardless of symptoms, SPTs were performed on most children.
attending the research center using 14 common food and aeroallergens (ALK-Albello, Horsholm, Denmark) [111]. Allergic sensitization was defined by having a positive SPT to at least one test with mean wheal diameter of 3 mm greater than the negative control. Since allergic sensitization is a dynamic phenotype, we used the concurrent status and thus allowed the risk to change over time.

4.3.3 FLG GENOTYPING

Blood and/or saliva samples were collected at ages 10 and/or 18 years from which genomic DNA was isolated. DNA samples were interrogated using GoldenGate Genotyping Assays on the BeadXpress Veracode platform (Illumina, Inc, SanDiego, CA) per Illumina’s protocol. Individuals carrying the minor allele for at least one of the FLG variants R501X, 2282del4, or S3247X were classified as having filaggrin haploinsufficiency. Detailed information on genotyping is provided by Ziyab et al [79].

4.3.4 STATISTICAL ANALYSIS

The 12-month period prevalence of eczema, asthma, rhinitis, and their comorbidities combinations was determined for each follow-up. Combinations of three disorders resulted in eight non-overlapping comorbidities groups (Table 4.1). To estimate the single and combined effects of the risk factors, FLG variants and repeated measures of allergic sensitization, on allergic disorders, generalized estimating equations (GEE) were used when estimating the covariance matrix to account for correlated observations and the within-child effect of the repeated measurements [88]. Risk ratios (RR) and their 95% confidence intervals (95% CIs) were estimated by applying log-binomial regression
models using the GENMOD procedure in SAS 9.3 (SAS, Cary, NC, USA). The “no allergic disorder” category formed the reference category. The main effects of FLG variants and allergic sensitization on the risk for single allergic disorders and comorbidities were estimated separately (single risk factor models). In a full model, the two main plus their interaction effect (FLG variants × allergic sensitization) were estimated. We repeated the aforementioned analysis while excluding the 1- or 2 years follow-up data from the GEE analysis to estimate whether possible misclassifications of phenotypes at early life biased our results. In the case of a possibly statistically significant interaction ($P_{\text{interaction}} < 0.1$), the combined effect was estimated as follows: $RR = \exp[(\beta_1 \times \text{allergic sensitization}) + (\beta_2 \times FLG \text{ variants}) + (\beta_3 \times [\text{allergic sensitization} \times \text{FLG variants}])]$. The term ‘combined effect’ was used to describe the joint impact of two individual risk factors plus their interaction on the occurrence of the outcome. In all GEE models, sex and age at follow-up were included as potential confounders. To evaluate statistical significance of main effects of covariates (not interaction terms), a $P$ value $\leq 0.05$ was used as indicator of statistical significance.

Population attributable fractions (PAFs), defined as the proportion of disease that can be related to a specific risk factor(s) [89], were estimated using the following formula: $PAF = p_e \times [(RR - 1)/RR]$, where $p_e$ is the proportion of diseased-individuals that are exposed [89, 90]. This approach yields valid PAF estimates in the presence of confounding and/or effect modification, only when using adjusted RRs [89, 90]. In case of statistically significant interactions between allergic sensitization and FLG variants, the RRs for the combined effect were used to estimate PAFs associated with both risk factors. If no interaction was detected, adjusted RRs were used to estimate separate PAFs.
for each of the two risk factors and an overall PAF was obtained using the following formula: $PAF_{overall} = 1 - [(1 - PAF_{allergic sensitization}) \times (1 - PAF_{FLG \ variants})]$ [89].

4.4 RESULTS

A total of 1,377, 1,210, 1,345, and 1,298 children, out of 1,456, had available information on eczema, asthma, and rhinitis diagnosis at ages 1-or-2, 4, 10, and 18 years, respectively (Table 4.1). SPT results were available for 515 (age 1-or-2: only children with any symptoms), 982, 1,036, and 853 children at ages 1-or-2, 4, 10, and 18 years, respectively. The proportion of positive SPTs increased from 20.6% to 41.4% between ages 1-or-2 and 18 years. Although SPT at age 1-or-2 was performed only on symptomatic children, the proportion of sensitized children at ages 1-or-2 and 4 years were similar (20.6% vs. 19.6%). The combined proportion of carriers (heterozygous) of FLG variants R501X, 2282del4 and S3247X was 10.3% (118/1150; Table 4.1).

4.4.1 PREVALENCE OF ALLERGIC DISORDERS

The period prevalence of “eczema only” was between 7.9% and 8.5% during the first 10 years of life and decreased to 5.6% at 18 years of age (Table 4.1). “Asthma only” was most prevalent (10.2%) at 4 years. The period prevalence of “rhinitis only”, despite the drop at age 4, demonstrated an increasing trend over time by reaching 20.4% at 18 years of age. The comorbidity of “eczema and asthma” and “eczema and rhinitis” ranged from 0.6% to 2.4% and 0.6% to 3.6%, respectively, between 1-or-2 and 18 years. Having “asthma and rhinitis” was most common (9.0%) at age 18 years. A coexistence of eczema, asthma, and rhinitis was found in approximately 2% over the study period.
4.4.2 Association Analysis: Single Risk Factor Models

*FLG* variants showed increased risk of “eczema only” (RR = 1.37, *P* = 0.101) and “rhinitis only” (RR = 1.27, *P* = 0.064; Table 4.2) that was not statistically significance, during the first 18 years of life. However, *FLG* variants predisposed to increased risk of having “eczema + asthma” (RR = 2.41, *P* < 0.01), “asthma + rhinitis” (RR = 1.52, *P* < 0.01), and a trend for increased risk of “eczema + rhinitis” (RR = 1.69, *P* = 0.077). Moreover, the risk of “eczema, asthma, and rhinitis” was 3.43 fold-higher among those with *FLG* haploinsufficiency than those with functional *FLG* gene. Accordingly, *FLG* variants showed stronger associations with allergic comorbidities than single disorders.

Allergic sensitization, statistically significantly, increased the risk of “eczema-“, “asthma-“, and “rhinitis-only” by 1.70-, 2.50-, 2.76-fold over the study period (Table 4.2). In addition, allergic sensitization predisposed to increased risk of having “eczema + asthma” (RR = 6.64, *P* < 0.01), “eczema + rhinitis” (RR = 3.48, *P* < 0.01), and “asthma + rhinitis” (RR = 4.71, *P* < 0.01). In particular, allergic sensitization was associated with an increased risk for the coexistence of “eczema, asthma, and rhinitis” (RR = 9.22, *P* < 0.01).

4.4.3 Association Analysis: Two Risk Factors Models

Simultaneously adjusting for *FLG* variants and allergic sensitization plus their interaction effect allowed us to determine their individual and combined effects. In these adjusted models, *FLG* variants did not show any association with allergic disorders (Table 4.3). In contrast, allergic sensitization was significantly associated with all allergic disorders. *FLG* variants and allergic sensitization showed possible interactive effects for
three of the studied comorbidities. Combined effect of allergic sensitization and FLG variants increased the risk of “eczema and asthma” (RR = 13.67, \( P < 0.01 \)), “asthma and rhinitis” (RR = 7.46, \( P < 0.01 \)), “eczema, asthma, and rhinitis” (RR = 23.44, \( P < 0.01 \); Table 4.3). To estimate whether the possible misclassifications of rhinitis/asthma at age 1-or-2 years influenced our results, we repeated the previous analyses while excluding data from the 1-or-2 years. Comparable results were obtained when we repeated our analysis by only including data from 4, 10, and 18 years follow-ups in the GEE analysis (data not shown).

4.4.4 Sex Differences

To determine whether sex is a potential effect modifier, we ran separate models for boys and girls. The effects of FLG variants and allergic sensitization on the risk of having single and multiple morbidities were similar for boys and girls. For instance, FLG variants associated with 2.95-fold (95% CI: 1.35 – 6.42, \( P < 0.01 \)) higher risk of having ‘eczema + asthma + rhinitis’ among boys; whereas, a 3.97-fold (95% CI: 1.96 – 8.05, \( P < 0.01 \)) higher risk was observed among girls (\( P_{\text{interaction for sex}} = 0.597 \); detailed data not shown).

4.4.5 Population Attributable Fractions (PAFs)

PAFs related to FLG variants and allergic sensitizations were estimated for the different allergic disorders using adjusted RR from Table 4.3. A larger fraction of allergic disorders in the population can be related to allergic sensitization than to FLG variants, when simultaneously adjusting for their effects (Table 4.4). Around 13.7% to
29.5% of single allergic disorders might be related to having both allergic sensitization and \(FLG\) variants. High PAFs for having “eczema, asthma, and rhinitis” (49%), “eczema + asthma” (33.7%), and “asthma + rhinitis” (21.2) were related to the combined effect of both allergic sensitization and \(FLG\) variants.

4.5 Discussion

In this well-characterized longitudinal study, allergic comorbidities were prevalent at all ages. For the first time, we assessed the single and combined effects of \(FLG\) variants and allergic sensitization on the development of single allergic disorders and comorbidities. \(FLG\) variants, when adjusting for the effect of allergic sensitization, did not show any association with allergic disorders. Against that, allergic sensitization, adjusting for the effect of \(FLG\) variants, increased the risk of all allergic disorders. Substantial combined effects of both factors were detected for allergic comorbidities. For instance, a 23-fold increased risk for having “eczema, asthma, and rhinitis” (approximately 2% of the cohort) was associated with the presence of both \(FLG\) variants and allergic sensitization.

Investigating and understanding the role of deviated immune responses, i.e. allergic sensitization, in the atopic diathesis have dominated the research field. However, the discovery of \(FLG\) variants and their pronounced role in the development of allergic disorders, eczema in particular, has caused a shift in the research paradigm toward understanding the contribution of dysfunctional skin barrier. \(FLG\) variants are considered to result in a compromised epidermal barrier that is characterized by increased penetration of allergens and microbes and elevated transepidermal water loss [60]. In
eczema, the role of FLG variants is well established; however, since FLG is not expressed in the respiratory epithelia its contribution to the development of allergic disorders of the airways is not clear [56, 60]. A suggested pathway is that cutaneous sensitization, facilitated by FLG variants, could lead to local and systematic inflammation at distant organs (i.e., lung and nasal tissues) [56]. Also the contrary is of importance, since it has been shown that inflammatory Th2 cytokines can also alter the integrity of the skin barrier, even in the absence of genetic variants that disrupt its integrity and function [112]. Hence, the two competing views: “inside-outside” which suggests that immunologic abnormality is the initiating and triggering step in the pathogenesis of allergic disorders and the “outside-inside” which posits that a defective epidermal barrier is the driver with epiphenomenal immunological sequelae, seem to be plausible and to some extent interdependent [77]. Therefore, simultaneously considering the contribution of genetic predisposition toward a defective skin barrier (i.e., FLG variants) and the propensity for immune dysregulation (i.e., allergic sensitization) could provide new and improved insights into the etiopathogenesis of allergic disorders.

Previous studies showed that FLG variants increased the risk of eczema and predisposed to higher risk of asthma and rhinitis only in the presence of coexisting eczema [59, 66]. Hence, suggesting that FLG variants increase the risk of allergic comorbidities, such as ‘asthma plus eczema’ and ‘rhinitis plus eczema’. Our results further support and add to the previously reported findings. However, we showed that FLG variants did not pose any increased risk of single and multiple allergic disorders while controlling for the effect of allergic sensitization. But, significant interactive effects of both risk factors were associated with increased risk of allergic comorbidities.
Although previous investigations inconsistently showed that FLG variants increased the risk of allergic sensitization [65, 66, 99], we did only observe a weak association between FLG variants and allergic sensitization in this cohort [79]. The possibility of an association between FLG variants and allergic sensitization cannot be totally excluded; however, since only around 15% of those with allergic sensitization carry FLG variants in our cohort, we speculate that any possible indirect effect of FLG variants on allergic diseases that goes through allergic sensitization to be minimal. Moreover, the possibility of non-independence between the two factors, also known as gene-environment correlation, might bias the interactive effects [113]. Due to the weak non-significant association between FLG variants and allergic sensitization in our cohort, we have little concern that the unlikely non-independence of the two factors might bias the observed interactive effects. In support of our analytical model, we clearly demonstrated in a previous report that FLG variants posed no increased risk of eczema in the absences of allergic sensitization, whereas, allergic sensitization increased the risk of eczema independent of FLG variants and the combined effect of both factors led to elevated risk above and beyond their individual effects [79]. Similarly, Wang et al. showed that the association between FLG variants and eczema was limited to a subset of children with IgE levels ≥ 100 kU/L [114]. Hence, we suggest considering the combined effect of both risk factors rather than taking the cause-effect approach could lead to a better risk assessment.

Our estimated PAFs suggested that approximately 21% to 49% of allergic comorbidities in the population may be attributed to the presence of both FLG variants and allergic sensitization. In agreement with results of this report, it has been shown that
a large proportion (38%) of allergic comorbidities can be explained by allergic sensitization alone [115]. Although, the contribution of FLG variants to the overall PAFs seems to be limited while controlling for the main effect of allergic sensitization, the PAFs associated with the combined effects of both risk factors (i.e., in the presence of possible interactive effects between FLG variants and allergic sensitization) is substantial. Our results suggest that FLG variants need allergic sensitization as a necessary component to execute its adverse effect. Therefore, early identification of those who are genetically predisposed to a dysfunctional skin barrier and who have increased allergic sensitization propensity may expedite the success of intervention/prevention efforts and measures.

The etiologic approach taken so far by research in the field of FLG variants and allergic disorders has substantially improved our understanding of these multifactorial disorders. However, as suggested by Ballardini et al., allergic disorders should not be studied as single entities [109], rather investigating their coexistence could assess in identifying shared underlying risk factors. Hence, we considered all possible and non-overlapping combinations of eczema, asthma, and rhinitis. Through this approach we were able to show that FLG variants increased the risk of allergic comorbidities rather than single disorders, suggesting its contribution towards a more severe and complex clinical phenotype. The effect of FLG variants when combined with the effect of allergic sensitization predisposed to even higher risk of allergic comorbidities. In contrast to investigating the coexistence of allergic disorders, evidence supporting the progression of events (i.e., allergic march) also exists [116, 117]. For instance, a valid assumption is that eczema predisposes to allergic sensitization, which in turn predicts asthma and rhinitis.
However, evidence supporting both pathways exists: (i) eczema precedes allergic sensitization and (ii) allergic sensitization precedes eczema [118, 119]. In the context of FLG variants, we have demonstrated, previously, that the combined effect of FLG variants and allergic sensitization predisposed to increased risk of subsequent eczema [79]. Also, we showed that the combination of FLG variants and eczema (FLG variants alone does not) increased the risk of subsequent allergic sensitization, which was limited to the first 10 years of life. Taken together, these observations suggest that multiple pathways could underlie the development of allergic disorders. Therefore, the development of allergic disorders is more complex and do not necessarily follow the atopic march paradigm. Although our approach does not take into account the sequence of the individual allergic manifestations, results of this report demonstrated that the co-manifestation of allergic disorders is common and FLG variants and allergic sensitization are common risk factors.

Major strengths of our study are the repeated phenotyping and the low attrition (ranging from 5% to 17%). FLG variants genotypes were available for the majority (80%) of the study population. Previously, we demonstrated that the genotyped sample did not differ from the total cohort with respect to multiple characteristics [79]. Although skin prick tests were performed at the different follow-ups to determine sensitization status, not measuring specific IgE is a limitation to our study. Moreover, since SPTs at ages 1 and 2 years were performed on symptomatic children, we speculate that selection bias leading to underestimated RRs might have been induced. Although proportions of SPT positivity at ages 1-or-2 years (20.6%) and 4 years (19.6%; SPT performed irrespective of symptoms) were similar, RRs associating allergic sensitization at age 1-or-
2 years with outcomes (i.e., eczema, asthma, and rhinitis) at the same age tended to be smaller than RRs relating allergic sensitization at age 4 years with outcomes at the same age (data not shown), since also the reference group had some symptoms. Hence, the estimated RRs at age 1-or-2 might be biased; however, the extent of the possible selection bias is minimal since our results did not noticeably change when we excluded the 1-or-2 years follow-up data from the analyses. Such observations further demonstrate the robustness of the GEE method in providing population averaged estimates (RRs) while accounting for the within-subject correlations of repeated measurements.

Misclassification of eczema cases is minimal since a high proportion of subjects showed typical manifestation of eczema in usual locations (antecubital or popliteal fossae, ankles, face or neck for 97% at 1 year, 91% at 2 years, 75% at 4 years, 86% at 10 years and 76% at 18 years) [25]. In addition, the prevalence of eczema in our cohort (13.7% at age 10 years and 12.3% at age 18 years) is comparable to results from ISAAC studies conducted among children aged 13–14 years in the United Kingdom (UK) (14.7% phase one and 10.6% phase three) and other studies conducted in the UK [5, 120, 121].

Other, potential limitations are the definition of asthma and rhinitis symptoms in early life. Our asthma conclusion at ages 10 and 18 years followed the ISAAC criteria [4], which was at that time not available for assessments at ages 1, 2, and 4 years. Although slightly different methods were used to define asthma; however, in a previous report we demonstrated that the minor change in asthma definition did not influence the validity of our asthma classification [37]. In addition, since it is difficult to differentiate between infectious and other forms of rhinitis in infancy, the elevated prevalence at 1-or-2 years (6.5% vs. 2.5% at age 4 years) might have been influenced by misclassifying ‘viral induced infectious
rhinitis’ [33]. The influence of the possible misclassification of rhinitis at 1-or-2 years on the overall results of the study is minimal since similar results were obtained when we reran our analyses while excluding the 1-or-2 year data.

In conclusion, comorbidities of eczema, asthma, and rhinitis are prevalent and both allergic sensitization and FLG variants are common risk factors that predispose to the comorbidity of allergic disorders. The combination of both risk factors was associated with elevated risk of coexisting allergic manifestations. Approximately 21% to 49% of the comorbidities in the population are related to both allergic sensitization and FLG variants. The coexistence of allergic disorders might reflect more severe clinical phenotypes that require more and longer medical care. However, investigations of comorbidities and their underlying etiological risk factors are limited. Therefore, additional etiological investigations regarding allergic comorbidities are recommended, as this will allow for a better phenotypic characterization.

4.6 ACKNOWLEDGMENTS

We thank participants and their families who have helped us with this project over the last two decades. We would like to acknowledge the help of all the staff at The David Hide Asthma and Allergy Research Centre in undertaking the 18 year and previous assessments of 1989 Isle of Wight birth cohort. The 18 year assessment of the IOW birth cohort and future analyses were funded by grants from the National Institute of Health, USA (R01 HL082925 and R01AI091905).
Table 4.1. Characteristics of the Isle of Wight study population at different ages

<table>
<thead>
<tr>
<th></th>
<th>Follow-up at age</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1-or-2 years</td>
</tr>
<tr>
<td>% (n)</td>
<td>n = 1377</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>50.8 (700)</td>
</tr>
<tr>
<td>No allergic disorders</td>
<td>68.9 (949)</td>
</tr>
<tr>
<td>Eczema only</td>
<td>8.5 (117)</td>
</tr>
<tr>
<td>Asthma only</td>
<td>4.9 (68)</td>
</tr>
<tr>
<td>Rhinitis only</td>
<td>6.5 (89)</td>
</tr>
<tr>
<td>Eczema + asthma</td>
<td>1.9 (26)</td>
</tr>
<tr>
<td>Eczema + rhinitis</td>
<td>1.8 (25)</td>
</tr>
<tr>
<td>Asthma + rhinitis</td>
<td>5.5 (75)</td>
</tr>
<tr>
<td>Eczema + asthma + rhinitis</td>
<td>2.0 (28)</td>
</tr>
<tr>
<td>Allergic sensitization</td>
<td>n = 515</td>
</tr>
<tr>
<td>≥ 1 positive SPT</td>
<td>20.6 (106)</td>
</tr>
<tr>
<td>FLG variants*</td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td></td>
<td>n = 1150</td>
</tr>
<tr>
<td>FLG variants</td>
<td>10.3 (118)</td>
</tr>
</tbody>
</table>

SPT: Skin prick test

* FLG variants were present in 118 participants of the 1150 who were genotyped. The prevalence of FLG variants changes based on the analytical sample.
Table 4.2. Single risk factor models: associations of FLG variants and allergic sensitization with allergic conditions: longitudinal analysis from age 1 or 2 to 18 years

<table>
<thead>
<tr>
<th>FLG variants genotype</th>
<th>Single risk factor model: FLG variants</th>
<th>Single risk factor model: Allergic sensitization</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>WT (k = 3831)</td>
<td>LOF (k = 442)</td>
</tr>
<tr>
<td>No allergic disorders (Ref.), % (k)</td>
<td>64.2 (2459)</td>
<td>54.3 (240)</td>
</tr>
<tr>
<td>Eczema only, % (k)</td>
<td>7.6 (291)</td>
<td>8.8 (39)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.37 (0.94 – 1.99)</td>
</tr>
<tr>
<td>Asthma only, % (k)</td>
<td>6.6 (253)</td>
<td>6.1 (27)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.07 (0.71 – 1.61)</td>
</tr>
<tr>
<td>Rhinitis only, % (k)</td>
<td>11.2 (428)</td>
<td>12.4 (55)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.27 (0.99 – 1.64)</td>
</tr>
<tr>
<td>Eczema + asthma, % (k)</td>
<td>1.4 (52)</td>
<td>2.9 (13)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>2.41 (1.28 – 4.57)</td>
</tr>
<tr>
<td>Eczema + rhinitis, % (k)</td>
<td>2.0 (76)</td>
<td>2.7 (12)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.69 (0.95 – 3.02)</td>
</tr>
<tr>
<td>Asthma + rhinitis, % (k)</td>
<td>5.4 (205)</td>
<td>6.8 (30)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.52 (1.04 – 2.23)</td>
</tr>
<tr>
<td>Eczema + asthma + rhinitis, % (k)</td>
<td>1.8 (67)</td>
<td>5.9 (26)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>3.43 (2.02 – 5.83)</td>
</tr>
</tbody>
</table>

WT: Wild-type; LOF: Loss-of-function; SPT –: Negative skin prick test; SPT +: Positive skin prick test; Ref.: represent the common reference category; k: refers to the total number of repeated measurements.

* In all GEE models sex and age at follow-up were included as potential confounders.

$ The presented numbers in parenthesis (k = ) refer to the number of repeated measurements. See Table 4.1 for more information on the number of individuals.

† $P$-value < 0.05

‡ $P$-value < 0.01
Table 4.3. Two risk factors model: associations of FLG variants and allergic sensitization with allergic conditions: longitudinal analysis from age 1-or-2 to 18 years

<table>
<thead>
<tr>
<th>FLG variants genotype</th>
<th>Allergic sensitization</th>
<th>RR_{Interaction} (P_{Interaction})</th>
<th>Combined effect$^\dagger$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>WT (k = 2707)$^¥$</td>
<td>LOF (k = 328)$^¥$</td>
<td>SPT – (k = 2164)$^¥$</td>
</tr>
<tr>
<td>No allergic disorders (Ref.), % (k)</td>
<td>56.9 (1540)</td>
<td>46.3 (152)</td>
<td>66.1 (1431)</td>
</tr>
<tr>
<td>Eczema only, % (k)</td>
<td>9.3 (252)</td>
<td>10.1 (33)</td>
<td>9.2 (200)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.11 (0.81 – 1.54)</td>
<td>1.00</td>
</tr>
<tr>
<td>Asthma only, % (k)</td>
<td>7.6 (205)</td>
<td>6.4 (21)</td>
<td>6.7 (144)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>0.99 (0.65 – 1.53)</td>
<td>1.00</td>
</tr>
<tr>
<td>Rhinitis only, % (k)</td>
<td>13.3 (360)</td>
<td>14.6 (48)</td>
<td>10.4 (226)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.21 (0.95 – 1.55)</td>
<td>1.00</td>
</tr>
<tr>
<td>Eczema + asthma, % (k)</td>
<td>1.6 (44)</td>
<td>4.0 (13)</td>
<td>1.0 (22)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>0.49 (0.08 – 3.03)</td>
<td>1.00</td>
</tr>
<tr>
<td>Eczema + rhinitis, % (k)</td>
<td>2.5 (68)</td>
<td>3.4 (11)</td>
<td>2.1 (46)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.52 (0.85 – 2.69)</td>
<td>1.00</td>
</tr>
<tr>
<td>Asthma + rhinitis, % (k)</td>
<td>6.7 (182)</td>
<td>8.2 (27)</td>
<td>3.4 (73)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>0.79 (0.34 – 1.89)</td>
<td>1.00</td>
</tr>
<tr>
<td>Eczema + asthma + rhinitis, % (k)</td>
<td>2.1 (56)</td>
<td>7.0 (23)</td>
<td>1.0 (22)</td>
</tr>
<tr>
<td>RR (95% CI)</td>
<td>1.00</td>
<td>1.02 (0.25 – 4.16)</td>
<td>1.00</td>
</tr>
</tbody>
</table>

WT: Wild-type; LOF: Loss-of-function; SPT –: Negative skin prick test; SPT +: Positive skin prick test; Ref.: represent the common reference category; k: refers to the total number of repeated measurements; RR_{Interaction}: Risk ratio associated with the interaction term.

* $P$-value < 0.1 was used to indicate presence of possible statistical interaction. If there was no evidence of interaction, the interaction term was removed from the regression model and only the main effects were modeled.

$^¥$ The presented numbers in parenthesis (k = ) refer to the number of repeated measurements. See Table 4.1 for more information on the number of individuals.

$^\dagger$ Combined effect was estimated only in the presence of possibly statistically significant interaction term (FLG variants × allergic sensitization; $P_{Interaction}$ < 0.1).

$^\ddagger$ $P$-value < 0.01.
**Table 4.4.** Population attributable fractions of different allergic conditions related to FLG variants and allergic sensitization

<table>
<thead>
<tr>
<th>FLG variants</th>
<th>Allergic sensitization</th>
<th>Overall PAF† (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>PAF (%)</td>
<td>PAF (%)</td>
</tr>
<tr>
<td>Eczema only</td>
<td>1.2</td>
<td>12.7</td>
</tr>
<tr>
<td>Asthma only</td>
<td>n/a⁵</td>
<td>22.6</td>
</tr>
<tr>
<td>Rhinitis only</td>
<td>2.1</td>
<td>28.0</td>
</tr>
<tr>
<td>Eczema + asthma</td>
<td>n/a⁶</td>
<td>n/a⁶</td>
</tr>
<tr>
<td>Eczema + rhinitis</td>
<td>4.8</td>
<td>28.7</td>
</tr>
<tr>
<td>Asthma + rhinitis</td>
<td>n/a⁶</td>
<td>n/a⁶</td>
</tr>
<tr>
<td>Eczema + asthma + rhinitis</td>
<td>n/a⁶</td>
<td>n/a⁶</td>
</tr>
</tbody>
</table>

PAF: population attributable fraction; n/a: not available

* PAFs were estimated using adjusted risk ratios obtained from Table 4.3.
† In the presence of statistically significant interaction, overall PAF were estimated using the RR for the combined effect obtained from Table 4.3.
‡ PAF was not estimated due to RR < 1.00.
§ PAF was not estimated due to the presence of possible statistical interaction (see Table 4.3).
¶ This value represents the PAF associated with allergic sensitization only since PAF associated with FLG variants was not estimable.
CHAPTER 5

RESULTS III

MANUSCRIPT # 3: DNA METHYLATION OF THE EPIDERMAL DIFFERENTIATION COMPLEX IN RELATION TO ECZEMA STATUS: AN EXPLORATORY STUDY

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5.1 **Abstract**

**Background:** Genetic variants within genes located across the epidermal differentiation complex (EDC) genomic region have shown associations with eczema risk. However, knowledge on the role of epigenetic mechanisms (e.g., DNA methylation) within the EDC locus on eczema risk is lacking.

**Objective:** To evaluate associations between DNA methylation of CpG sites across the EDC genomic region with eczema status.

**Methods:** DNA methylation levels of 256 CpG sites spanning the EDC locus were measured using Illumina's 450K array in three semi-independent cohorts; a subsample of the Isle of Wight birth cohort (n = 367 participants aged 18 years, *discovery cohort*), *replication cohort I* (n = 146; parents of the F2 generation), and *replication cohort II* (n = 94; F2 generation). Associations between eczema status and DNA methylation were first assessed in the discovery cohort using linear regression. Methylation sites showing an association with eczema in the discovery cohort at a false discovery rate p-value < 0.1 were taken forward for replication.

**Results:** Differential DNA methylation of CpG site cg12048339 (located within promoter of *S100A6* gene) was associated with eczema specifically among female participants of all study cohorts; whereas, aberrant DNA methylation of cg10959711 (located within promoter of *S100A11* gene) associated with eczema among male participants in all study samples.

**Conclusion:** This exploratory study demonstrated that DNA methylation of the EDC genomic region could be an important factor in the development of eczema in a sex-specific manner. Future studies corroborating our findings are needed.
5.2 Introduction

The elevated life-time prevalence of eczema among children (up to 30%) and adults (up to 10%) makes eczema a global public-health burden [22]. Immunologic dysregulation and defective epidermal barrier are the main hallmarks of eczema [46]. In last decades, etiologic research have focused on immune-related factors, including genetics of immune system, in the pathogenesis of eczema; however, the discovery, in 2006, of loss-of-function variants in the filaggrin gene (FLG) have caused a shift in the research paradigm towards understanding factors that regulate the formation and integrity of the epidermal barrier [53, 68]. Filaggrin haploinsufficiency, defined as partial or complete loss of filaggrin (filament-aggregating protein) protein, is associated with the development of an impaired epidermal barrier that is characterized by increased penetration of environmental substances and elevated transepidermal water loss [93]. As yet, FLG variants are the strongest and most replicated genetic risk factors for eczema development [59].

The FLG gene is located within a dense cluster of genes encoding structural and regulatory proteins that are essential for keratinocyte differentiation, known as the epidermal differentiation complex (EDC) [76, 81, 82]. The EDC locus, spanning 2 Mb on human chromosomal region 1q21, harbors around 60 genes encoding loricrin and involucrin proteins and gene-family clusters encoding: (i) small proline-rich (SPRR) and late cornified envelope (LCE) proteins, (ii) calcium-binding proteins (S100A), and (iii) S100-fused type proteins (SFTP) [76, 81, 82]. Apart from FLG variants, a limited number of studies identified variants in other genes within the EDC that are associated with eczema. For instance, conflicting results as to whether polymorphisms in the hornerin
gene (HRNR), located 78 kb from FLG gene, are associated with eczema risk have been reported [122, 123]. Regarding SPRR3, Marenholz et al. showed that a 24-bp deletion in the gene was associated with eczema [124]. However, Stemmler et al. did not find evidence of association between eczema and other EDC genetic variants than FLG variants [125]. Both, genome-wide linkage and association studies have detected signals of linkage/association between eczema and the EDC genomic locus [122, 126-128]. Morar et al. showed that the genetic linkage of eczema to the EDC region (LOD score = 3.57) was not totally diminished after accounting for FLG variants (LOD score = 2.03) [129]. Hence, this finding point towards the presence of other genetic variants within the EDC that influence eczema development. On the other hand, the limited success, apart from FLG variants, of associating EDC gene variants with eczema is not promising. Moreover, candidate-gene and genome-wide association studies have only explained a small proportion of the high heritability (up to 85%) and clustering of eczema within families [68, 69].

Epigenetic mechanisms (e.g., DNA methylation) may offer a complementary explanation when considering the role of EDC on eczema. DNA methylation addresses heritable genetic regulatory elements that alter gene activity (e.g., expression) without changing the underlying nucleotide sequence of the DNA [130] have been speculated as plausible predisposing factors in the development of eczema and might as well account for the unexplained genetic effects [69]. Currently, there is a lack of knowledge on the role of epigenetic mechanisms, in particular DNA methylation, of the EDC genomic locus in relation to eczema risk. Therefore, a comprehensive DNA methylation profiling of the EDC genomic region could further add understanding for this genomic region in
development of eczema. To this end, in an EDC-wide screening this explorative study aimed to evaluate the association of DNA methylation across the EDC region with eczema status in participants of the Isle of Wight birth cohort (aged 18 years, F1 generation). The analyses further test any significant findings identified in the screening (discovery) in two semi-independent samples (pregnant cohort participant, their partners, and their offspring, the F2 generation).

5.3 Materials and Methods

5.3.1 Participants and Characteristics

The Isle of Wight birth (IOW) cohort, an unselected population-based study, was recruited between January 1989 and February 1990 in the Isle of Wight, UK, to study the natural history and etiology of allergic diseases. After exclusion of adoptions, perinatal deaths, and refusal for follow-up, 1,456 children (95%) were enrolled, with follow-up assessments conducted at 1, 2, 4, 10, and 18 years of age. Ethics approvals were obtained from the Isle of Wight Local Research Ethics Committee at recruitment and for the subsequent follow-ups (06/Q1701/34). Written informed consent was obtained from parents, participants, or both. In this analysis, the discovery cohort is based on a subsample (n = 367, aged 18 years) of the IOW cohort that was randomly selected for epigenetic study (Figure 5.1). We consider two cohorts to test the reproducibility of the screening. Replication cohort I (n = 146; parents of the F2 generation) includes participants of the IOW cohort (28.8% were part of the discovery cohort) plus new participants. Expecting mothers and their partners were assessed before delivery. Replication cohort II (n = 94; F2 generation) includes the F2 newborns of the IOW
cohort participants (Figure 5.1). Infants were followed-up at ages 3-, 6-, and 12-months. In all study samples, eczema was defined as chronic or chronically relapsing, itchy dermatitis lasting more than 6 weeks with characteristic morphology and distribution, following Hanifin and Rajka criteria [84].

5.3.2 DNA METHYLATION PROFILING

For the discovery cohort, DNA was extracted from whole blood collected at age 18 years using a standard salting out procedure [85], and bisulfite-treated for cytosine to thymine conversion using the EZ 96-DNA methylation kit (Zymo Research, CA, USA). Similarly, whole blood of replication cohort I participants’ and cord blood of replication cohort II participants’ were collected and DNA was extracted and bisulfite converted following the aforementioned standard procedures. In all samples, Genome-wide DNA methylation was assessed using the Illumina Infinium HumanMethylation450 BeadChip (Illumina, Inc., CA, USA), which interrogates > 484,000 CpG sites associated with approximately 24,000 genes. For the purpose of this report, we restricted our analysis to 256 CpG sites that span the EDC genomic region (location on human chromosome 1: 151958685 – 153628983). Arrays were processed using a standard protocol [86], with multiple identical control samples assigned to each bisulfite conversion batch to assess assay variability and samples randomly distributed on microarrays to control against batch effects [131]. The BeadChips were scanned using a BeadStation, and the methylation level calculated for each queried CpG locus (cytosine-guanine dinucleotides) using the Methylation Module of BeadStudio software. We used the DNA methylation profiles of our participants to predict their cell-type composition via a method proposed
by Houseman et al. [132] and validated by Koestler et al. [133]. This method identified CpG loci from our data which were within differentially methylated regions (DMRs) known to distinguish between white blood cell types, then utilized our Beta-values to predict the proportions of CD8+ T-cells, CD4+ T-cells, natural killer cells, B-cells, monocytes and granulocytes for each sample.

5.3.3 Statistical Analysis

After performing quality control measures on DNA methylation data, Beta-values presented as the proportion of methylated (M) over methylated (M) and unmethylated (U) sites \((\text{Beta-value} = M/[c+M+U])\), with \(c\) being constant to prevent dividing by zero) were calculated for each CpG site [87]. Beta-values provide intuitive biological interpretation (i.e., % methylation); however, (i) being constrained between values of 0 (unmethylated) and 1 (completely methylated) and (ii) demonstrating high heteroscedasticity in the lower and upper ends of the methylation range has raised concerns on the validity of this measure when performing statistical analysis [91]. The conversion of Beta-values to \(M\)-values \((M\text{-value} = \log_2(Beta\text{-value}/[1 – Beta\text{-value}]))\) has been shown to overcome the limitations of beta-values [91]. Moreover, the use of \(M\)-values, when performing statistical analysis, demonstrated increased reliability in terms of detecting differentially methylated CpG sites; however, \(M\)-values cannot be directly interpreted [91, 92]. Therefore, following previous recommendation [91], we used \(M\)-values when performing the statistical analysis and reported the statistics at the scale of Beta-values.

All statistical analyses were conducted using SAS® version 9.3 (SAS Institute, Cary, NC, USA). In the discovery cohort, linear regression was used to test the
association between DNA methylation \((M\text{-values})\) of all CpG sites spanning the EDC region with eczema status adjusting for sex and cell-type composition. To minimize false-positive findings identified in 256 models, CpG sites with a false discovery rate (FDR) adjusted p-value < 0.1 were selected and considered as potentially associated with eczema. The selected CpG sites, in the discovery cohort, were taken forward and tested for their possible association with eczema status in replication cohorts I and II. Then, the selected CpG sites were tested for association with eczema in males and females separately.

5.4 RESULTS

The 12-month period prevalence of eczema was 12.6%, 15.6%, and 16.7% in the discovery cohort, replication cohort I, and replication cohort II, respectively (Table 5.1). The period prevalence of eczema was statistically significantly higher among women compared to men in the discovery cohort (15.2% vs. 7.4%, p-value = 0.034) and replication cohort I (21.1% vs. 5.9%, p-value = 0.017). In contrast, eczema was more common among infant boys (21.1%) in comparison with infant girls (11.8%) in replication cohort II.

Epigenome-wide DNA methylation profiling, covering more than 484,000 CpG sites across the genome, was performed in all three study samples. We restricted our analysis to 256 CpG sites that spanned the EDC genomic region. In the discovery cohort, screening for possible associations between DNA methylation of CpG sites within the EDC and eczema identified five CpG sites that are differentially methylated between eczema-affected and eczema-free participants with FDR adjusted p-values < 0.1 (Table
The five identified CpG sites were then tested in women and men participants separately. The sex-specific results showed that DNA methylation of the five CpG sites was statistically significantly associated with eczema in women but not in men (Table 5.2). However, the directions of the effect sizes of three out of the five CpG sites (cg10959711, cg13493250, and cg01910639) were in agreement between women and men. For instance, DNA methylation at cg13493250 locus (located within the LCE1F gene promoter region) was, on average, 4% higher in women and by 3% in men, when comparing eczema-affect to eczema-free participants.

The five identified CpG sites were further tested for association with eczema in replication cohort I (Table 5.3). None of the CpG sites showed statistically significant association with eczema. The direction of the effect size of two CpG sites (cg10959711 and cg12048339) was in agreement when comparing results of the discovery cohort (Table 5.2) with results of replication cohort I (Table 5.3). The sex-stratified analyses showed DNA methylation of two CpG sites in females (cg10959711 and cg12048339) and four CpG sites in males (cg10959711, cg13493250, cg03969260, and cg12048339) were in agreement, in regard to effect size direction, between the discovery cohort and replication cohort I.

Finally, associations between the five CpG sites and eczema were tested in the replication cohort II (Table 5.4). In regard to statistical significance, none of the associations were replicated; however, two CpG sites (cg10959711 and cg12048339) showed similar magnitude of association as seen in the discovery cohort. In infant girls, DNA methylation of one CpG site (cg12048339) demonstrated similar effect across the discovery cohort and replication cohort II. In infant boys, DNA methylation of two CpG
sites (cg10959711 and cg01910639) showed similar effects across the *discovery cohort* and *replication cohort II*.

In the analysis that considered both sexes, there were two CpG sites (cg10959711 and cg12048339) that showed agreement in respect to the magnitude of their effect sizes across the three study samples. Furthermore, inspecting the sex-specific analyses showed that one CpG site (cg12048339) among female participants and one CpG site among male participants (cg10959711) demonstrated similar effect sizes across the three study samples. For instance, DNA methylation of cg12048339 was 2% (*discovery cohort* and *replication cohort I*) and 1% (*replication cohort II*) lower in eczema-affected as compared to eczema-free female participants. On the other hand, DNA methylation of cg10959711 locus was 1% (*discovery cohort* and *replication cohort I*) and 3% (*replication cohort II*) higher in eczema-affected as compared to eczema-free male participants.

5.5 DISCUSSION

This explorative study, for the first time, investigated the possible association between DNA methylation across the EDC genomic region with eczema in three semi-independent population based samples. In the *discovery cohort*, DNA methylation of five CpG sites was associated with eczema status. Although results of replicating these associations did not demonstrate statistical significance, two CpG sites (cg10959711 and cg12048339) showed similar effect sizes across the three study samples. Noticeably, DNA methylation of CpG site cg12048339 was associated with eczema specifically among female participants of all study cohorts; whereas, DNA methylation of
cg10959711 associated with eczema among male participants in all study samples. Such results might point towards sex-specific associations between DNA methylation across the EDC locus and eczema. Moreover, DNA methylation within the EDC genomic region might explain some of the genome-wide linkage and association signals that relate the EDC locus with eczema.

Both of the identified sex-specific CpG sites are located within genes that belong to the *S100A* gene-family cluster on the EDC locus. The cg12048339 is located within the promoter region of *S100A6* gene, specifically 1500 base-pairs upstream of transcription start site (TSS1500). Similarly, the cg10959711 is located within the TSS1500 genomic region of *S100A11* gene. The *S100A* genes, overall, share similar gene structure by containing two introns and three exons (exon-2 and exon-3 are translated) and encode proteins that, in general, contain two calcium-binding EF-hand motifs [134, 135]. The S100A proteins are involved in various biological functions and processes, such as cell cycle and differentiation, transcription, motility, and inflammation [134]. The clustering of *S100A* genes within the EDC locus, in addition to evidence supporting their involvement in epidermal differentiation [135], suggests a biologically-plausible role in the pathogenesis of skin-related diseases. For instance, elevated serum levels of S100A8 and S100A9 proteins have been shown to be related to abnormal differentiation of keratinocytes in skin of psoriasis patients [136].

The current state of knowledge suggests that increased methylation of gene promoters is associated with reduced genes expression; whereas, decreased methylation is associated with increased gene expression; however, emerging evidence suggest that such a regulatory role highly depends on the context in which methylation occurs in the
genome and is not limited to methylation of promoter regions [137, 138]. In this study we observed that among female participants with eczema as compared to those without eczema, DNA methylation of cg12048339 (TSS1500 of S100A11 gene) was reduced, on average, by 1% to 2% across the three study samples. In contrast, DNA methylation of cg10959711 (TSS1500 of S100A6 gene) among males with eczema as compared to eczema-free participants was increased, on average, by 1% to 3% across the study cohorts. Such results suggest that altered methylation of CpG sites within the EDC locus could serve as markers of eczema in a sex-specific manner.

Existing evidence support the regulatory role of DNA methylation on the activity of S100A genes [139]. Although we observed associations between DNA methylation in two CpG sites with eczema, results of this preliminary report should be interpreted with caution. First, DNA methylation, in this report, was not measured in the respective tissue of the disease (i.e., skin); therefore, the extent to which DNA methylation measured in peripheral blood (or cord blood) relate to other tissues and whether can be used as a biomarker for phenotype variation is unclear and an area of current scientific dispute [137, 140]. Second, the discrepancy, apart from the two identified CpG sites, in the results across the three study samples could be due to (i) the differential phenotype expression over age and/or (ii) the dynamic nature of DNA methylation that changes with respect to age and environmental exposures. For instance, previously we observed a sex-reversal in the prevalence of eczema during adolescence period with more girls developing eczema and more boys outgrowing their eczema [25], such a sex-switchover in the risk could be mediated by changes in DNA methylation.
There is no doubt that this study can only serve as a starting point for future studies due to the limited sample sizes of the studied cohorts. The *discovery cohorts* is the largest sample (n = 367) that we studied and from which we obtained the initial significant findings. *Replications cohorts I and II* were smaller in size (n = 146 and n = 94, respectively) and failed to replicate, in terms of statistical significance, results of the *discovery cohort*. The sex-specific analyses were based on even smaller groups. This said, we acknowledge the fact that two out of five CpG sites were in agreement in regard to their effect sizes across the three study cohorts. Moreover, the observed modest effect sizes are not unusual, apart from cancer studies, for studies investigating complex diseases. For example, a 2.07% difference in DNA methylation of the *ORMDL3* gene when comparing asthmatic (87.90%) to non-asthmatic (85.83%) children has been reported [141]. Currently, there is no consensus as to whether modest changes in DNA methylation are as important or less important when compared to larger changes in regard to the development of complex diseases. To this end, previously we demonstrated that an interaction between genetic variants in the *FLG* gene with adjacent DNA methylation predisposed to greater risk of eczema [80]. These results indicated that the association between *FLG* loss-of-function variants and eczema is modulated by DNA methylation. Therefore, simultaneously considering the DNA methylation of the EDC locus and the underlying genetic variants might provide further insights into the genetics/epigenetics of eczema than when considering each factor separately. Taken together, future studies should incorporate DNA methylation, genetic predisposition, gene expression, and proteins levels to provide a better assessment of the risk.
In conclusion, this exploratory study demonstrated that DNA methylation of the EDC genomic region could be an important factor in the development of eczema and might as well explain some of the observed genome-wide linkage and association signals relating the EDC locus with eczema. Interestingly, we observed gender-specific associations between DNA methylation of CpG sites and eczema, which further support the fact that eczema development is different across the two sexes. Future studies confirming these preliminary observations are needed and should obtain DNA methylation profiles from the disease-tissue to increase reliability and relevant of any findings.
Figure 5.1. Flow diagram of the different analyzed study samples.
Table 5.1. Prevalence of eczema in the different study samples stratified by sex

<table>
<thead>
<tr>
<th>Study Sample</th>
<th>Total</th>
<th>Females</th>
<th>Males</th>
<th>p-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Discovery cohort (F1)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eczema at age 18 years, % (n/total)</td>
<td>12.6</td>
<td>15.2</td>
<td>7.4</td>
<td>0.034</td>
</tr>
<tr>
<td>Missing, n</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td><strong>Replication cohort I (F2 parents)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eczema, % (n/total)</td>
<td>15.6</td>
<td>21.1</td>
<td>5.9</td>
<td>0.017</td>
</tr>
<tr>
<td>Missing, n</td>
<td>5</td>
<td>3</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td><strong>Replication cohort II (F2 offspring)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eczema at age 1 year, % (n/total)</td>
<td>16.7</td>
<td>11.8</td>
<td>21.1</td>
<td>0.354</td>
</tr>
<tr>
<td>Missing, n</td>
<td>22</td>
<td>9</td>
<td>13</td>
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</tr>
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*P-value comparing proportions of females to males.
Table 5.2. Association of CpG sites in the EDC genomic region with eczema status: results of the discovery cohort

<table>
<thead>
<tr>
<th>Both sexes</th>
<th>Gene</th>
<th>Placement</th>
<th>Average methylation (5th – 95th percentile)</th>
<th>Effect size (SE)</th>
<th>Raw p-value</th>
<th>FDR adj. p-value</th>
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<td>No eczema</td>
<td></td>
<td></td>
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<tr>
<td>cg10959711</td>
<td>S100A6</td>
<td>TSS1500</td>
<td>0.11 (0.07 – 0.16)</td>
<td>0.12 (0.09 – 0.19)</td>
<td>0.02 (0.004)</td>
<td>0.00023 0.055</td>
</tr>
<tr>
<td>cg13493250</td>
<td>LCE1F</td>
<td>TSS1500</td>
<td>0.38 (0.29 – 0.49)</td>
<td>0.41 (0.31 – 0.52)</td>
<td>0.03 (0.010)</td>
<td>0.00052 0.055</td>
</tr>
<tr>
<td>cg03969260</td>
<td>SPRR2G</td>
<td>3’ UTR</td>
<td>0.93 (0.90 – 0.95)</td>
<td>0.94 (0.92 – 0.95)</td>
<td>0.01 (0.002)</td>
<td>0.00065 0.055</td>
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<tr>
<td>cg12048339</td>
<td>S100A11</td>
<td>TSS1500</td>
<td>0.67 (0.62 – 0.72)</td>
<td>0.65 (0.61 – 0.69)</td>
<td>-0.02 (0.005)</td>
<td>0.00118 0.071</td>
</tr>
<tr>
<td>cg01910639</td>
<td>S100A6</td>
<td>Body</td>
<td>0.73 (0.63 – 0.81)</td>
<td>0.75 (0.67 – 0.83)</td>
<td>0.03 (0.009)</td>
<td>0.00139 0.071</td>
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<table>
<thead>
<tr>
<th>Female participants</th>
<th></th>
<th></th>
<th>Yes eczema</th>
<th>No eczema</th>
<th></th>
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<tbody>
<tr>
<td>cg10959711</td>
<td>S100A6</td>
<td>TSS1500</td>
<td>0.11 (0.06 – 0.18)</td>
<td>0.12 (0.09 – 0.22)</td>
<td>0.02 (0.006)</td>
<td>0.0016</td>
</tr>
<tr>
<td>cg13493250</td>
<td>LCE1F</td>
<td>TSS1500</td>
<td>0.38 (0.29 – 0.50)</td>
<td>0.40 (0.31 – 0.52)</td>
<td>0.04 (0.011)</td>
<td>0.0012</td>
</tr>
<tr>
<td>cg03969260</td>
<td>SPRR2G</td>
<td>3’ UTR</td>
<td>0.93 (0.91 – 0.95)</td>
<td>0.93 (0.92 – 0.95)</td>
<td>0.01 (0.002)</td>
<td>0.0018</td>
</tr>
<tr>
<td>cg12048339</td>
<td>S100A11</td>
<td>TSS1500</td>
<td>0.67 (0.62 – 0.72)</td>
<td>0.65 (0.60 – 0.70)</td>
<td>-0.02 (0.005)</td>
<td>0.0002</td>
</tr>
<tr>
<td>cg01910639</td>
<td>S100A6</td>
<td>Body</td>
<td>0.74 (0.62 – 0.82)</td>
<td>0.75 (0.66 – 0.83)</td>
<td>0.03 (0.012)</td>
<td>0.0066</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Male participants</th>
<th></th>
<th></th>
<th>Yes eczema</th>
<th>No eczema</th>
<th></th>
<th></th>
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</thead>
<tbody>
<tr>
<td>cg10959711</td>
<td>S100A6</td>
<td>TSS1500</td>
<td>0.10 (0.08 – 0.13)</td>
<td>0.11 (0.09 – 0.16)</td>
<td>0.01 (0.005)</td>
<td>0.0251</td>
</tr>
<tr>
<td>cg13493250</td>
<td>LCE1F</td>
<td>TSS1500</td>
<td>0.39 (0.29 – 0.48)</td>
<td>0.42 (0.28 – 0.49)</td>
<td>0.03 (0.021)</td>
<td>0.1959</td>
</tr>
<tr>
<td>cg03969260</td>
<td>SPRR2G</td>
<td>3’ UTR</td>
<td>0.93 (0.91 – 0.94)</td>
<td>0.92 (0.90 – 0.94)</td>
<td>-0.01 (0.003)</td>
<td>0.9959</td>
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<td>cg12048339</td>
<td>S100A11</td>
<td>TSS1500</td>
<td>0.66 (0.62 – 0.71)</td>
<td>0.67 (0.64 – 0.69)</td>
<td>0.01 (0.011)</td>
<td>0.8144</td>
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<td>cg01910639</td>
<td>S100A6</td>
<td>Body</td>
<td>0.71 (0.65 – 0.75)</td>
<td>0.73 (0.67 – 0.76)</td>
<td>0.02 (0.012)</td>
<td>0.2849</td>
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</tbody>
</table>

SE: Standard error; FDR: False discovery rate; Adj.: adjusted

Effect size is calculated as average methylation difference between eczema-affected and eczema-free participants using linear regression models. Effect sizes of the ‘both sexes’ models were adjusted for sex and cell-type composition.
<table>
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<tr>
<th>CpG ID</th>
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<th>Placement</th>
<th>Both sexes</th>
<th></th>
<th></th>
<th>Female participants</th>
<th></th>
<th></th>
<th>Male participants</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
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<td>0.10 (0.08 – 0.11)</td>
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<tr>
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<td>TSS1500</td>
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<td>0.38 (0.29 -0.51)</td>
<td>0.39 (0.31 – 0.51)</td>
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<tr>
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SE: Standard error; FDR: False discovery rate; Ajd.: adjusted
* Effect size is calculated as average methylation difference between eczema-affected and eczema-free participants using linear regression models. Effect sizes of the 'both sexes' models were adjusted for sex and cell-type composition.
Table 5.4. Association of CpG sites in the EDC genomic region with eczema status: results of the *replication cohort II*

<table>
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<th>Both sexes</th>
<th>Gene Placement</th>
<th>Average methylation (5th – 95th percentile)</th>
<th>Effect size (SE)</th>
<th>Raw p-value</th>
</tr>
</thead>
<tbody>
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</tr>
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<td>0.85 (0.75 – 0.89)</td>
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</table>

<table>
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<td>0.85 (0.77 – 0.89)</td>
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<table>
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<th>Raw p-value</th>
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</tr>
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<td>cg12048339</td>
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SE: Standard error; FDR: False discovery rate; Adj.: adjusted

Effect size is calculated as average methylation difference between eczema-affected and eczema-free participants using linear regression models. Effect sizes of the ‘both sexes’ models were adjusted for sex and cell-type composition.
CHAPTER 6

DISCUSSION

6.1 SECTION STRUCTURE

In this section I will tie together the major findings of this dissertation and their relevant interpretation. In general, results of chapters 3 and 4 will by synthesized together as they were (i) based on the same study sample and (ii) investigated the role of FLG variants and allergic sensitization on the development of single (chapter 3) and multiple (chapter 4) allergic diseases. In contrast, results of chapter 5 will be discussed separately since it was based on three semi-independent study samples and investigated the potential role of DNA methylation of the EDC genomic locus on the development of eczema.

6.2 OVERVIEW OF FINDINGS

The first two research-related chapters (i.e., chapters 3 and 4) of this dissertation aimed at elucidating the role of FLG variants and allergic sensitization on the development of allergic diseases. Chapter 3 focused on the possible effect modification by allergic sensitization and/or eczema on the association between “FLG variants and asthma” and “FLG variants and rhinitis”. I demonstrated that both allergic sensitization and eczema status are possible effect modifiers of the association between FLG variants and asthma. In contrast, neither eczema nor allergic sensitization modified the association of FLG variants with rhinitis in a statistically significant manner. In chapter 4, I showed that the coexistence of eczema, asthma, and rhinitis is prevalent and allergic sensitization
and FLG variants jointly predisposed to increased risk of allergic comorbidities. On the other hand, chapter 5 investigated the role of DNA methylation across the EDC genomic region on the risk of eczema. Results of this exploratory study demonstrated possible gender-specific associations between DNA methylation of CpG sites and eczema in the discovery study sample; however, the results, across the three semi-independent study samples, did not identify consistent pattern of associations between DNA methylation of CpG sites in EDC locus with eczema.

6.3 FLG VARIANTS AND ALLERGIC SENSITIZATION IN RELATION TO ALLERGIC DISEASES

Studies investigating the natural history (long-term prognosis) of allergic diseases have improved our understanding of the biological mechanisms that underlie the clinical manifestation of these complex disorders. For instance, in previous reports we have demonstrated that a sex-switchover in the prevalence of eczema [25], asthma [37], and rhinitis [33] occurs around puberty, with more adolescent girls being affect. Etiologic investigations have indicated the importance of environmental, immunologic, and genetic factors in the pathogenesis of allergic diseases [23, 26, 110, 142-144]. Moreover, close and complex inter-relatedness between the manifestations of eczema, asthma, and rhinitis have been demonstrated [45, 48, 103]; however, common mechanisms and pathways underlying their development is an area of ongoing scientific research. The ‘allergic march’ concept suggests that allergic disorders develop in a sequential pattern, starting with eczema in early infancy and later in childhood developing asthma and rhinitis [41, 106]. However, supporting replicable evidence and consensus is far from complete [108, 145, 146]. An opposing concept is the “coexistence” of allergic morbidities [42, 45, 109,
Therefore, identifying common risk factors that predispose to the development of single and/or multiple allergic diseases is of importance and will further elucidate the etiology of such complex diseases.

Allergic sensitization (deviated immune responses) has been widely associated with different allergic diseases [48, 49, 96]. Similarly, FLG variants (representing defective epidermal barrier) have recently emerged as strong independent risk factors for eczema [59]. Furthermore, FLG variants increase the risk of asthma and rhinitis only in the presence of coexisting eczema [59, 60]. Apart from eczema, it has been suggested that FLG variants predispose to allergic disorders of the airways by facilitating the penetration of allergens through the defective epidermal barrier, which in turn might lead to local and systematic inflammation at distant organs (e.g., lung and nasal tissues) [56, 147]. In this dissertation I have focused on FLG variants and allergic sensitization as they have been considered as common risk factors that predispose and link the manifestation of eczema, asthma, and rhinitis [49, 60, 79]. That said, our current state of knowledge lacks clear understanding on their single and joint role in the development of single (chapter 3) and multiple (chapter 4) allergic diseases. Utilizing the prospective nature of the Isle of Wight birth cohort study, I tested if FLG variants interact with allergic sensitization and/or eczema on the risk of later asthma and rhinitis. Specifically, I investigated as to whether interplay between “FLG variants and allergic sensitization” and/or “FLG variants and eczema” predisposed to the development of subsequent asthma and rhinitis. Next, I tested whether FLG variants and allergic sensitization jointly predisposed to the coexistence of eczema, asthma, and rhinitis.
6.3.1 Single Allergic Diseases

Thus far, FLG variants are the strongest and most replicated genetic risk factors for eczema development [59, 66]. In a previous report, we have corroborated the reported association between FLG variants and eczema and further demonstrated that allergic sensitization is an important effect modifier of this association [79]. We showed that FLG variants increased the risk of eczema only in the presence of allergic sensitization and the combined effect of both risk factors was associated with elevated risk of eczema (RR = 2.92, 95% CI: 1.47–5.77) that was above and beyond their individual effects [79]. In this dissertation, I sought to explore the possibility that allergic sensitization could also modify the reported associations of FLG variants with asthma and rhinitis.

Prior studies have shown that FLG variants are associated with asthma only in the presence of coexisting eczema [59, 66]. However, this possible effect modification by eczema was not consistently demonstrated for the association between FLG variants and rhinitis [65, 66]. To this end, I have tested the hypothesis that allergic sensitization, rather than eczema status, modifies the association of FLG variants with asthma and rhinitis. I have tested my hypothesis under two models: (i) concurrent effect models in which both the exposure and the outcome coexisted at the same time and (ii) delayed effect models in which I used the preceding cumulative allergic sensitization status (or eczema status), i.e. all SPTs results (or eczema status) from assessments that are prior to the outcome assessment, when testing the interaction between FLG variants and allergic sensitization (or eczema) on the risk of subsequent asthma/rhinitis.

In stratified analyses (concurrent model), both eczema and allergic sensitization status modulated the association between FLG variants and asthma. Statistical
interactions on multiplicative scale were tested to determine whether the observed heterogenous effects were statistically significantly different. A statistically significant interaction was found only between \(FLG\) variants and allergic sensitization on the risk of asthma. In delayed effect models, which take the time order of risk factors and disease occurrences into account, I have tested the interaction of ‘\(FLG\) variants with preceding allergic sensitization’ and ‘\(FLG\) variants with preceding eczema’ on the risk of subsequent asthma. Results of delayed effect models showed that \(FLG\) variants in interaction with preceding allergic sensitization and preceding eczema increased the risk of subsequent asthma. Hence, indicating that both allergic sensitization and eczema might act as effect modifiers for the association between \(FLG\) variants and asthma.

In regard to rhinitis, results of stratified analyses (concurrent model) demonstrated that eczema status did not modify the association between \(FLG\) variants and rhinitis; however, this association was modified when stratifying by allergic sensitization status. But, there was no evidence for an interactive effect (statistical interaction) neither between \(FLG\) variants and eczema nor between \(FLG\) variants and allergic sensitization on the risk for rhinitis. Similarly, results of delayed effect models showed that neither preceding eczema nor preceding allergic sensitization interacted with \(FLG\) variants on the risk of subsequent rhinitis.

These findings indicate that \(FLG\) variants may need eczema and/or allergic sensitization to execute their adverse effects on asthma. The finding of an interaction between \(FLG\) variants and allergic sensitization in this study improves our understanding of the possible link between genetics of the epidermal barrier (i.e., \(FLG\) variants) and a respiratory disorder (i.e., asthma). Such an observation further supports the hypothesis
that cutaneous sensitization priming, partially facilitated by FLG variants, could lead to local and systematic inflammation at distant organs (e.g., lung) [56, 98]. Our results of an interactive effect between FLG variants and allergic sensitization on the risk of asthma and the lack of such interaction on the risk of rhinitis suggest that the pathway through which FLG variants predispose to asthma and rhinitis may be different. For instance, the expression of filaggrin protein was not detected in the human bronchial epithelium; however, it has been shown that filaggrin is expressed in the nasal vestibule [64, 94]. That said, the extent to which filaggrin expression in the nasal vestibular lining influence the development of rhinitis is yet to be investigated by future studies.

In summary, the first part of this dissertation has demonstrated that (i) FLG variants are associated with single allergic diseases such as asthma and rhinitis and (ii) allergic sensitization and eczema status act as possible effect modifiers of the association between FLG variants and asthma. The combined effects of ‘preceding allergic sensitization and FLG variants’ and ‘preceding eczema and FLG variants’ increased the risk of subsequent asthma by 4.93-fold and 3.33-fold, respectively, during the study period (i.e., the first 18 years of life). In contrast, the association between FLG variants and rhinitis was not modulated by either allergic sensitization or eczema status. Taken together, our results demonstrated differential etiological pathways underlying the development of asthma and rhinitis and future studies corroborating our observations are needed.
6.3.2 Multiple Allergic Diseases

After exploring the role of allergic sensitization and FLG variants on the development of single allergic diseases, I investigated their single and joint contribution towards the development of allergic comorbidities (i.e., coexistence of eczema, asthma, and rhinitis). First, I constructed eight non-overlapping combinations of the three allergic disorders (Table 2.1), which were used in subsequent analyses for this part of the dissertation.

Although previous studies have demonstrated the importance of allergic sensitization and FLG variants on the development of single allergic diseases [49, 60, 96], no prior study have investigated the joint role of both risk factors on the development of allergic comorbidities. In this dissertation, I have demonstrated that the coexistence of “eczema, asthma, and rhinitis” affect around 2% of the study population. Both risk factors, when tested separately, were associated with single and multiple allergic diseases. However, FLG variants, when adjusting for the effect of allergic sensitization, did not show any association with allergic disorders. Remarkably, interactions between the two risk factors were associated with substantial combined effects on the risk of allergic comorbidities. For instance, a 23-fold increased risk for having “eczema, asthma, and rhinitis” was associated with the presence of both FLG variants and allergic sensitization. Moreover, we estimated that approximately 21% to 49% of allergic comorbidities in the population may be attributed to the presence of both FLG variants and allergic sensitization.

In summary, results of this part of the dissertation suggests that comorbidities of eczema, asthma, and rhinitis are prevalent and both allergic sensitization and FLG
variants are common risk factors that predispose to the comorbidity of allergic disorders. The combination of both risk factors was associated with elevated risk of coexisting allergic manifestations. Although in the previous section of this dissertation I showed that \(FLG\) variants and allergic sensitization predispose to single allergic diseases, in this section I further demonstrated the importance of their joint role on the co-manifestation of allergic diseases. Therefore, simultaneously considering the contribution of genetic predisposition toward a defective skin barrier (i.e., \(FLG\) variants) and the propensity for immune dysregulation (i.e., allergic sensitization) provided new and improved insights into the pathogenesis of allergic disorders.

6.3.3 Strengths and Limitations

The first two research-related chapters (i.e., chapters 3 and 4) in this dissertation were based on information collected in the Isle of Wight birth cohort study. Major strengths of our prospective study, covering the first 18 years of life, are the repeated phenotyping, objective assessments of allergic sensitization status, and the low loss to follow-up (ranged from 5% to 17%). Moreover, the majority (80%) of the study participants were genotyped for \(FLG\) variants common among populations of European ancestry. We showed, previously, that the genotyped study participants did not differ from the total cohort with regard to multiple characteristics [79]. Hence, there is no indication of selection bias that could pose a threat to the validity of our study. Misclassification of eczema cases is minimal since a high proportion of subjects showed typical manifestation of eczema in the usual locations (antecubital or popliteal fossae,
ankles, face or neck for 97% at 1 year, 91% at 2 years, 75% at 4 years, 86% at 10 years and 76% at 18 years) [25].

Potential limitations are the definition of asthma and rhinitis symptoms in early life. Our asthma conclusion at ages 10 and 18 years followed the ISAAC criteria [4], which was at that time not available for assessments at age 1, 2, and 4 years. Although slightly different methods were used to define asthma, in a previous report we have shown that the minor change in asthma definition over time did not influence the validity of our asthma classification (i.e., no noticeable change in asthma prevalence up to 10 years of age) [37]. In addition, since it is difficult to differentiate between infectious and other forms of rhinitis in infancy, the elevated prevalence at 1- or-2 years might have been influenced by misclassifying ‘viral induced infectious rhinitis’ [33]. However, by applying repeated measurements analysis based solely on assessments later in childhood and adolescence (4, 10, and 18 years), we believe that the influence of the possible misclassification of rhinitis at 1- or-2 years on the overall results of the study is minimal, since similar results were obtained. Moreover, since SPTs at age 1- or-2 years were performed on symptomatic children, we speculate that selection bias leading to underestimated RRs might have been induced. Although proportions of SPT positivity at ages 1- or-2 years (20.6%) and 4 years (19.6%; SPT performed irrespective of symptoms) were similar, RRs associating allergic sensitization at age 1- or-2 years with outcomes (i.e., eczema, asthma, and rhinitis) at the same age tended to be smaller than RRs relating allergic sensitization at age 4 years with outcomes at the same age, since also the reference group had some symptoms. Hence, the estimated RRs at age 1- or-2 might be underestimated; however, the extent of the possible selection bias is minimal since our
results did not noticeably change when we excluded the 1-or-2 years follow-up data from the analyses. Such observations further demonstrate the robustness of the GEE method in providing population averaged estimates (RRs) while accounting for the within-subject correlations of repeated measurements.

6.4 DNA METHYLATION OF THE EDC AND ECZEMA RISK

The third research-related chapter (i.e., chapter 5) of this dissertation focused on the role of DNA methylation across the EDC genomic region in relation to eczema status. In this explorative study, I have screened for associations between DNA methylation of 256 CpG sites across the EDC locus with eczema status at age 18 years (discovery cohort), and subsequently replicated the significant findings in two semi-independent cohorts. Results of this analysis showed that DNA methylation of two CpG sites (cg10959711 and cg12048339) demonstrated similar effect sizes across the three study samples. Noticeably, DNA methylation of CpG site cg12048339 was associated with eczema specifically among female participants of all study cohorts; whereas, DNA methylation of cg10959711 associated with eczema among male participants in all study samples. Such results might point towards sex-specific associations between DNA methylation across the EDC locus and eczema.

Both of the identified sex-specific CpG sites are located within genes that belong to the $S100A$ gene-family cluster on the EDC locus. The cg12048339 is located within the promoter region of $S100A6$ gene, specifically 1500 base-pairs upstream of transcription start site (TSS1500). Similarly, the cg10959711 is located within the TSS1500 genomic region of $S100A11$ gene. The $S100A$ genes, overall, share similar gene
structure by containing two introns and three exons (exon-2 and exon-3 are translated) and encode proteins that, in general, contain two calcium-binding EF-hand motifs [134, 135]. The S100A proteins are involved in various biological functions and processes, such as cell cycle and differentiation, transcription, motility, and inflammation [134]. The clustering of S100A genes within the EDC locus, in addition to evidence supporting their involvement in epidermal differentiation [135], suggests a biologically-plausible role in the pathogenesis of skin-related diseases. In this study we observed that among female participants with eczema as compared to those without eczema, DNA methylation of cg12048339 (TSS1500 of S100A11 gene) was reduced, on average, by 1% to 2% across the three study samples. In contrast, DNA methylation of cg10959711 (TSS1500 of S100A6 gene) among males with eczema as compared to eczema-free participants was increased, on average, by 1% to 3% across the study cohorts. Such results suggest that altered methylation of CpG sites within the EDC locus could serve as markers of eczema in a sex-specific manner.

6.4.1 STRENGTHS AND LIMITATIONS

There is no doubt that this study can only serve as a starting point for future studies due to the limited sample sizes of the studied cohorts. The discovery cohorts is the largest sample (n = 367) that we studied and from which we obtained the initial significant findings. Replications cohorts I and II were smaller in size (n = 146 and n = 94, respectively) and failed to replicate, in terms of statistical significance, results of the discovery cohort. The sex-specific analyses were based on even smaller groups. Moreover, DNA methylation, in this report, was not measured in the respective tissue of
the disease (i.e., skin); therefore, the extent to which DNA methylation measured in peripheral blood (or cord blood) relate to other tissues and whether can be used as a biomarker for phenotype variation is unclear and an area of current scientific dispute [137, 140]. The discrepancy, apart from the two identified CpG sites, in the results across the three study samples could be due to (i) the differential phenotype expression over age and/or (ii) the dynamic nature of DNA methylation that changes with respect to age and environmental exposures. For instance, previously we observed a sex-reversal in the prevalence of eczema during adolescence period with more girls developing eczema and more boys outgrowing their eczema [25], such a sex-switchover in the risk could be mediated by changes in DNA methylation.

6.5 Conclusion

In this dissertation I have investigated the role of FLG variants (representing an impaired epidermal barrier) and allergic sensitizations (representing deviated immune responses) on the development of single and multiple allergic diseases. Results of this dissertation clearly demonstrated that FLG variants and allergic sensitization are common factors that predispose and link the manifestation of different allergic diseases (i.e., eczema, asthma, and rhinitis). First, I have shown that allergic sensitization and eczema modulated the association between FLG variants and asthma, but not rhinitis. Hence, implying that the mechanisms and pathways through which FLG variants predispose to increased risk of asthma and rhinitis may be different. Second, a coexistence of allergic disorders was demonstrated to be frequent; and allergic sensitization and FLG variants jointly increased risk of allergic comorbidities. The coexistence of allergic diseases may
represent more severe and complex clinical phenotypes that require more and longer medical care. The interactive effect and the elevated proportion of allergic comorbidities associated with allergic sensitization and FLG variants emphasize their joint importance in the pathogenesis of allergic disorders. Third, DNA methylation across the EDC genomic region might influence the risk of eczema in a sex-specific manner.

In summary, this dissertation provided insights on (i) the joint role of FLG variants and allergic sensitization on the development of single and multiple allergic diseases and (ii) the role of DNA methylation of the EDC locus on the development of eczema. Therefore, results of this dissertation helped improve our ability to better identify and stratify those who share similar risk characteristics, which may also enhance clinical management and the development of efficient intervention and prevention strategies. Future studies corroborating our findings of the joint role of FLG variants and allergic sensitization on the development of single and multiple allergic diseases are needed. Moreover, future studies confirming our preliminary observations of sex-specific associations between DNA methylation and eczema are needed and should obtain DNA methylation profiles from the disease-tissue (i.e., skin) to increase reliability and enhance relevance of any findings.
REFERENCES


10. Asher MI. Recent perspectives on global epidemiology of asthma in childhood. *Allergol Immunopathol (Madr)* 2010; **38**:83-7.


42. Sun HL, Yeh CJ, Ku MS, Lue KH. Coexistence of allergic diseases: patterns and frequencies. *Allergy Asthma Proc* 2012; **33**:e1-4.


51. Weidinger S, Baurecht H, Naumann A, Novak N. Genome-wide association studies on IgE regulation: are genetics of IgE also genetics of atopic disease? *Curr Opin Allergy Clin Immunol* 2010; **10**:408-17.


104. Reed CE. The natural history of asthma. *J Allergy Clin Immunol* 2006; **118**:543-8; quiz 49-50.


118. Almqvist C, Li Q, Britton WJ et al. Early predictors for developing allergic disease and asthma: examining separate steps in the 'allergic march'. *Clin Exp Allergy* 2007; **37**:1296-302.


139. Lesniak W. Epigenetic regulation of S100 protein expression. *Clin Epigenetics* 2011; **2**:77-83.


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