Biogenic Nitrogen Gas Production at the Oxic–Anoxic Interface in the Cariaco Basin, Venezuela

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Abstract. Excess nitrogen gas (N\textsubscript{x}2\textsuperscript{s}) was measured in samples collected at six locations in the eastern and western sub-basins of the Cariaco Basin, Venezuela, in September 2008 (non-upwelling conditions) and March 2009 (upwelling conditions). During both sampling periods, N\textsubscript{x}2\textsuperscript{s} concentrations were below detection in surface waters, increasing to \( \sim 22\, \mu\text{mol N kg}^{-1} \) at the oxic–anoxic interface ([O\textsubscript{2}] < \( \sim 4\, \mu\text{mol kg}^{-1}, \sim 250 \, \text{m} \)). Below the oxic–anoxic interface (300–400 m), the average concentration of N\textsubscript{x}2\textsuperscript{s} was 24.7 \( \pm 1.9\, \mu\text{mol N kg}^{-1} \) in September 2008 and 27.5 \( \pm 2.0\, \mu\text{mol N kg}^{-1} \) in March 2009, i.e., N\textsubscript{x}2\textsuperscript{s} concentrations within this depth interval were \( \sim 3\, \mu\text{mol N kg}^{-1} \) higher (\( p < 0.001 \)) during the upwelling season compared to the non-upwelling period. These results suggest that N-loss in the Cariaco Basin may vary seasonally in response to changes in the flux of sinking particulate organic matter. We attribute the increase in N\textsubscript{x}2\textsuperscript{s} concentrations, or N-loss, observed during upwelling to: (1) higher availability of fixed nitrogen derived from suspended and sinking particles at the oxic–anoxic interface and/or (2) enhanced ventilation at the oxic–anoxic interface during upwelling.

1 Introduction

Dissolved inorganic nitrogen (DIN or “fixed” nitrogen), includes nitrate, nitrite, and ammonium. This nutrient often limits primary production in marine systems (Codispoti, 1989; Smith, 1984) and thus plays a role in the modulation of climate (Altabet et al., 2002; Falkowski, 1997; Ganeshram et al., 2000; Gruber, 2004). In the ocean, new DIN is primarily derived from nitrogen fixation (N\textsubscript{2} \rightarrow NH\textsubscript{4}+) by microbial diazotrophs. The microbial processes that result in the conversion of DIN back to dinitrogen gas (N-loss) are the primary oceanic sinks for fixed nitrogen (Codispoti et al., 2001; Gruber and Sarmiento, 1997). Several studies suggest that today’s rate of ocean N-loss exceeds the rate of nitrogen fixation by over 200 Tg per year (Brandes and Devol, 2002; Codispoti, 2007). This estimate is based in part on the accumulation of nitrate deficit (with respect to the Redfield N:P ratio [N\textsuperscript{+}]; Gruber and Sarmiento, 1997) as a function of the residence time of oxygen-depleted
water masses (Gruber, 2004; Gruber and Sarmiento, 1997, 2002). Whether this imbalance in the nitrogen inventory is due to an underestimation of nitrogen fixation or an overestimation of N-loss is a topic of current debate (Capone and Knapp, 2007; Codispoti, 2007; Deutsch et al., 2007; Dalsgaard et al., 2012; DeVries et al., 2012). More recently, the removal of fixed nitrogen has been estimated by measuring excess \( N_2 \) (\( N_2^{xs} \), with respect atmospheric equilibrium), such as in the Arabian and Baltic Seas and the eastern tropical South Pacific (Chang et al., 2010; Devol et al., 2006; Löffler et al., 2011), in order to reduce uncertainties associated with regional variations in N : P ratios and the complexities of DIN conversion pathways. DeVries et al. (2012) used \( N_2^{xs} \) measurements coupled with a global circulation model to estimate oceanic water-column N-loss rates. In anoxic waters of the Black Sea, interannual variations in \( N_2^{xs} \) were attributed to changes in the downward flux of organic matter from phytoplankton blooms (Fuchsman et al., 2008). Despite these efforts many questions remain regarding the role of organic matter flux for fueling water column N-loss (Brettar and Rheinheimer, 1992; Liu and Kaplan, 1984; Ward et al., 2008).

The Cariaco Basin offers an opportunity to examine the linkage between surface productivity and the loss of fixed nitrogen at depth. This basin exhibits water column N-loss, high seasonal phytoplankton production (\( \sim 700 \) to \( 2300 \) mg C m\(^{-2} \) d\(^{-1} \)) and high vertical flux of particulate organic matter (\( \sim 40 \) to \( 100 \) mg C m\(^{-2} \) d\(^{-1} \); Muller-Karger et al., 2001; Scranton et al., 2006; Thunell et al., 2007; Zhang and Millero, 1993). One of the first measurements of marine \( N_2^{xs} \) was conducted in the anoxic waters of the Cariaco Basin by Richards and Benson (1961) using \( N_2 / Ar \) ratiometry. In this manuscript we build on Richards and Benson’s work, and report new estimates of biogenic \( N_2 \) gas production at several locations within the Cariaco Basin during upwelling and non-upwelling periods. We tested the hypothesis that biogenic production of \( N_2 \) varies both spatially and temporally within the anoxic waters of the basin in response to changes in the flux of organic matter. We also compared DIN deficits in the water column with \( N_2^{xs} \) measurements to understand whether the removal of fixed nitrogen by denitrification was balanced by the biogenic production of \( N_2 \). We assumed that heterotrophic (canonical water column denitrification) and chemautotrophic (anammox) consumption of DIN were the only sources of biogenic \( N_2 \) (Chang et al., 2010; Devol et al., 2006).

2 Study area

The Cariaco Basin is a 1400 m deep, tectonically derived depression on the continental shelf of Venezuela (Schubert, 1982; Fig. 1). Its sill (\( \sim 100 \) m) has two deeper channels, La Tortuga (135 m) and Centinela (145 m; Lídiz et al., 1969; Richards, 1975). The exchange of water between the Cariaco Basin and the Caribbean Sea/Atlantic Ocean is limited. Cariaco waters > 150 m deep are poorly ventilated; waters at the oxic–anoxic interface (\( \sim 250 \) m) have residence times of months to years, while bottom waters have residence times of years to decades (Holmén and Rooth, 1990; Zhang and Millero, 1993). The basin is subject to seasonal changes in the wind intensity along the coast related to the meridional migration of the Intertropical Convergence Zone, which drives alternating upwelling (January–May) and thermally stratified conditions (August–November) (Astor et al., 2003).

During upwelling, primary production (PP) and particulate organic matter (POM) vertical fluxes are high; the 14-yr PP and sediment trap particulate organic carbon (POC) flux (225 m) averages are 1871 \( \pm \) 394 and 80 \( \pm \) 20 mg C m\(^{-2} \) d\(^{-1} \), respectively (Muller-Karger et al., 2001, 2010). During the non-upwelling period, primary production and settling POC flux averages are significantly lower, 874 \( \pm \) 136 mg C m\(^{-2} \) d\(^{-1} \) and 60 \( \pm \) 20 mg C m\(^{-2} \) d\(^{-1} \), respectively.

The decomposition of the sinking POC contributes to present-day anoxic and sulfidic conditions below \( \sim 250 \) m (Scranton et al., 2001; Zhang and Millero, 1993).

3 Methods

3.1 Sample collection

Samples were collected from the R/V Hermano Ginés during two cruises to Cariaco Basin (1–6 September 2008 and 9–13 March 2009). A total of 36 stations were occupied for general oceanographic observations during each cruise, while dissolved \( N_2 \) samples were collected at six stations: three in the eastern sub-basin (stations 10, 11 and 13) and three in the western sub-basin (stations 21, 29 and 32) (Table 1, Fig. 1). These six stations all have water depths exceeding 300 m and included the oxic–anoxic interface. Hydrographic data were obtained using a Seabird® SBE-25 conductivity-temperature-depth (CTD) device equipped with an Eco chlorophyll fluorometer (Wetlabs), an SBE43 dissolved oxygen sensor, and a rosette with 12 Teflon-coated 8L Niskin bottles.

Stations 10 and 21 featured the deepest (>1300 m) profiles of anoxic waters. Station 10 is the location of the CARIACO Ocean Time Series Station (10.5°N, 64.6°W), sampled monthly since November 1995. Combined with stations 11 and 13, these three sites were chosen to help assess spatial changes in the eastern Cariaco Basin. Station 29 was sampled to investigate possible riverine influences on local hydrographic conditions, as it was closer to the mouth of the Tuy River, the largest river discharging into the Cariaco Basin. Finally, station 32 was selected to evaluate the effect of the influx of Caribbean waters through Centinela Channel (Fig. 1).
Table 1. Location, profile depth, and chlorophyll a concentration at the depth of chlorophyll maximum (DCM) at stations sampled for N\textsubscript{2}\textsuperscript{s} during September 2008 and March 2009 in the Cariaco Basin.

<table>
<thead>
<tr>
<th>Station #</th>
<th>Latitude (° N)</th>
<th>Longitude (° W)</th>
<th>Profile depth (m)</th>
<th>September 2008 [Chl a] (µg L\textsuperscript{-1})</th>
<th>DCM (m)</th>
<th>March 2009 [Chl a] (µg L\textsuperscript{-1})</th>
<th>DCM (m)</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>10.5</td>
<td>64.7</td>
<td>1300</td>
<td>0.31</td>
<td>50</td>
<td>0.60</td>
<td>20</td>
</tr>
<tr>
<td>11</td>
<td>10.7</td>
<td>64.7</td>
<td>400</td>
<td>0.61</td>
<td>57</td>
<td>0.60</td>
<td>33</td>
</tr>
<tr>
<td>13</td>
<td>10.5</td>
<td>65.0</td>
<td>400</td>
<td>0.63</td>
<td>61</td>
<td>0.54</td>
<td>42</td>
</tr>
<tr>
<td>21</td>
<td>10.7</td>
<td>65.7</td>
<td>1300</td>
<td>0.42</td>
<td>50</td>
<td>1.25</td>
<td>43</td>
</tr>
<tr>
<td>29</td>
<td>10.7</td>
<td>65.9</td>
<td>400</td>
<td>0.42</td>
<td>65</td>
<td>1.15</td>
<td>38</td>
</tr>
<tr>
<td>32</td>
<td>10.8</td>
<td>65.7</td>
<td>400</td>
<td>0.52</td>
<td>50</td>
<td>2.10</td>
<td>50</td>
</tr>
</tbody>
</table>

Fig. 1. Location of the Cariaco Basin and stations sampled for N\textsubscript{2}\textsuperscript{s} (circles) in September 2008 and March 2009. Station 10 corresponds to the CARIACO Ocean Time Series Station (10.5° N, 64.7° W). Broken lines trace the bathymetry of the basin. The black star shows the location of the Tuy River mouth.

Chlorophyll a and nutrient samples were collected at the 36 stations during each cruise, from the surface to 100 m (1, 20, 30, 50, 100 m); below this depth chlorophyll concentrations are typically below the detection limit of 0.01 µg L\textsuperscript{-1}. Chlorophyll a determinations were carried out using methanol extractions (see Muller-Karger et al., 2001) and a Turner Designs model 10 fluorometer as described by Falkowski and Kiefer (1985) and Holm-Hansen et al. (1965).

DIN and soluble reactive phosphorus (SRP) samples were collected at 1, 20, 50 and 100 m at stations 10, 11, 13 and 29, and at the surface at stations 21 and 32. Additional nitrate and ammonium samples were collected at 200, 250, 300 and 400 m. Nutrient data from the CARIACO Ocean Time Series Station (station 10 in Fig. 1) collected during September 2008 and March 2009 were used in this study as well (data available at the CARIACO Ocean Time Series website: http://www.imars.usf.edu/CAR/). Nutrient determinations were conducted at the University of South Florida following the protocols of Gordon et al. (2000) and Strickland and Parsons (1972). We used H\textsubscript{2}S measurements from samples collected on separate cruises at the CARIACO Station between 2007 and 2009 (April and November 2007, May 2008, and January and May 2009) since sulfide determinations were not conducted during our cruises. These sulfide data are presented in Li et al. (2008, 2010).
Water for the dissolved N\textsubscript{2} analyses was collected using Niskin bottles at 1, 50, 100, 200, 275, 300, and 400 m at the six stations selected for this study. At stations 10 and 21, additional samples were collected at 500, 750, and 1300 m. Samples were collected using 60 mL biological oxygen demand (BOD) bottles using the filling procedure for Winkler O\textsubscript{2} determinations. A 100 µL aliquot of saturated HgCl\textsubscript{2} was added to each sample for preservation. Bottles were sealed with glass stoppers, and standing seawater was left around the exterior of the glass stopper at the mouth of the bottle to inhibit gas exchange with the atmosphere. The mouth of the BOD bottles, including the stopper and standing water, were sealed with a plastic cap to avoid spilling of the standing seawater, and samples were kept in a cooler or refrigerator at \(\sim 4^\circ C\) until analyzed.

3.2 Gas analyses

High-precision estimates of nitrogen gas were conducted at the School of Marine Science and Technology, University of Massachusetts Dartmouth, using isotope ratio mass spectrometry (IRMS). Dissolved gases were extracted from each sample by pumping an aliquot at 10 mL min\(^{-1}\) through a chambered microfiber-microporous gas extractor coupled to a continuous flow of He carrier gas. The extracted gas was then passed through water, CO\textsubscript{2} and O\textsubscript{2} traps before transferring to a GV IsoPrime IRMS for simultaneous measurements of N\textsubscript{2} (mass 28) and Ar (mass 40). A liquid nitrogen cold trap was also used to remove H\textsubscript{2}S from samples collected within the sulfidic portion of the water column. Gas ratios from samples were compared against calibrated references of artificial compressed gas mixtures of N\textsubscript{2} and Ar covering the expected dissolved gas ratios. Further calibration also included comparing gas measurements in distilled water equilibrated with air at controlled temperatures of 10 and 20°C (\(\pm 0.01^\circ C\)) to theoretical determinations based on the solubility of these gases. The analytical precision of N\textsubscript{2}/Ar measurements was better than 0.3 %, and thus the limit of detection of N\textsubscript{2} was \(\sim 0.3 \mu mol kg^{-1}\).

N\textsubscript{2}\textsubscript{S} was calculated from deviations of the sample N\textsubscript{2}/Ar ratio from equilibrium values expected from in situ measurements of potential temperature and salinity (Hamme and Emerson, 2004). This largely eliminates artifacts related to changes in gas concentration caused by physical effects on the dissolution of N\textsubscript{2} due to variations in temperature and salinity. N\textsubscript{2}/Ar ratios were, in turn, normalized to ratios at atmospheric equilibrium (N\textsubscript{2} : Ar\textsubscript{sample} / N\textsubscript{2} : Ar\textsubscript{atm. equilibrium}) to detect departures of N\textsubscript{2} concentrations from expected equilibrium values due to biogenic N\textsubscript{2} production. N\textsubscript{2}\textsubscript{S} can also arise from physical processes in the near-surface region influencing air–sea gas exchange. Positive N\textsubscript{2}\textsubscript{S} is produced by air injection due to bubble collapse that can occur in remote water mass formation regions (Hamme and Emerson, 2002; Hamme and Severinghaus, 2007), whereas negative anomalies can arise locally by in situ heating when there is diminished exchange with the atmosphere.

Background (or abiotic) N\textsubscript{2}/Ar ratios in the Cariaco Basin were estimated based on data collected at the Bermuda Atlantic Time Series Station (BATS) (Nicholson et al., 2010; M. Altabet, unpublished data, 2012). Normalized N\textsubscript{2}/Ar ratios at BATS are near atmospheric equilibrium in surface waters (\(\sim 27^\circ C\)) and \(\sim 1.5 \%\) saturated at 1000 m (\(\sim 6^\circ C\) waters). Background N\textsubscript{2}/Ar ratios in the Cariaco Basin were then derived from the linear regression equation for the correlation between salinity-normalized temperature and measured N\textsubscript{2}/Ar ratios at BATS (\(y = -0.0232x + 1.0176; R^2 = 0.88\)). Our background N\textsubscript{2}/Ar ratio estimates in Cariaco are based upon the assumption that Subtropical Underwater (SUW), the major source water to the basin, has a similar vertical distribution of N\textsubscript{2}/Ar supersaturation to that observed at BATS since SUW is formed in the eastern subtropical North Atlantic.

4 Results

4.1 Hydrographic data

Profiles of dissolved oxygen (O\textsubscript{2}) concentration and density (\(\sigma_T\)) were measured at the six stations sampled for gas analyses during September 2008 and March 2009 (Fig. 2). For both seasons, oxygen concentrations decreased from \(\sim 185 \pm 12 \mu mol kg^{-1}\) at the surface to below detection at 300 m. There was higher [O\textsubscript{2}] at the \(\sim 25–26\) isopycnal surface (\(\sim 0–120 m\)) in March 2009 likely as a result of increased vertical mixing and ventilation; the [O\textsubscript{2}] at 25 isopycnal was \(\sim 35 \mu mol kg^{-1}\) higher in March 2009 than in September 2008. Sulfide was detected immediately below the depth at which oxygen was below detection ([H\textsubscript{2}S] > 1 \mu mol kg\(^{-1}\) at 270 m) between 2007 and 2009 and increased with depth to a mean maximum value of \(65 \pm 3 \mu mol kg^{-1}\) at 1300 m.

A plot of salinity (S) versus potential temperature (\(\theta\)) for the six stations selected for excess N\textsubscript{2} measurements during the two cruises indicates that water masses below \(\sim 150 m\) in the Cariaco Basin during the two sampling periods were similar (Fig. 3). During September 2008, stations in the western sub-basin (stations 21, 29 and 32) showed lower salinities near the surface than those in the eastern sub-basin (stations 10, 11 and 13). This surface water is derived from the western tropical North Atlantic and the Caribbean Sea. The salinity maximum, which is associated with SUW, shows a slight warming during the upwelling season when this water mass is closer to the surface and therefore exposed to sunlight.

4.2 Nutrients and chlorophyll a

Nutrient concentrations at the surface were nearly undetectable during both seasons due to biological uptake. They were slightly higher in the upper 50 m at the six stations...
Fig. 2. Profiles of mean [O$_2$], [H$_2$S] and water column temperature versus depth in the upper 400 m of the Cariaco Basin. Oxygen and temperature profiles are constructed from data collected at all six stations sampled during September 2008 and March 2009 for N$_x$s determinations. Average [H$_2$S] was calculated from data collected on separate cruises at the CARIACO Station between 2007 and 2009 (see Sect. 3.1). Mean oxygen concentrations are also plotted against density ($\sigma_T$).

Samples collected for gas determinations (Table 2) in March 2009 as a result of upwelling, compared to September 2008. Nitrite and SRP concentrations in the top 100 m were below 1 µmol kg$^{-1}$ in both seasons. A peak in [NO$_3^-$] of 10–12 µmol kg$^{-1}$ was observed around 200 m, and [NO$_3^-$] was below detection at 300 m. Ammonium concentrations were low but variable near the surface (<0.5 µmol kg$^{-1}$), with slightly higher values in the upper 50 m during March 2009 compared to September 2008. Ammonium concentrations showed a minimum at 200 m in both seasons. From 250 m to the bottom, [NH$_4^+$] increased at a rate of ~0.04 µmol kg$^{-1}$ m$^{-1}$ reaching values as high as ~23 µmol kg$^{-1}$ at 1300 m (stations 10 and 21; data not shown).

Samples collected at selected stations for N$_x$s measurements showed nutrient concentrations similar to those from the CARIACO Ocean Time Series during corresponding months (September 2008 and March 2009; Fig. 4). As expected, all nutrient measurements in the upper 100 m from the CARIACO Ocean Time Series showed higher concentrations in March 2009 (upwelling conditions) than in September 2008 (non-upwelling conditions). Nitrite showed a large peak at 100 m in September 2008, but a secondary peak near the oxic–anoxic interface was not detected during this time, possibly due to wide sample spacing. In March 2009, however, a sharp secondary nitrite maximum (0.86 µmol kg$^{-1}$) was measured at ~270 m. Nitrate, SRP and ammonium concentrations below 100 m showed similar vertical profiles during both periods.

A deep chlorophyll maximum (DCM) was observed at 50–65 m in September 2008 (0.48 ± 0.12 µg L$^{-1}$; Table 1), and at 20–50 m in March 2009 (1.04 ± 0.60 µg L$^{-1}$). This shoaling of the DCM is characteristic of the upwelling season (Astor et al., 2003). Chlorophyll $a$ concentrations at the DCM in the eastern sub-basin were on the order of 0.60 µg L$^{-1}$, while in the western sub-basin concentrations ranged 1.15-2.10 µg L$^{-1}$.
Table 2. Average nutrient concentrations from stations sampled for \(N_{2}^{ss}\) determinations during September 2008 and March 2009 in the Cariaco Basin. Values near the oxic–anoxic interface are shown in bold captions.

<table>
<thead>
<tr>
<th>Depth (m)</th>
<th>([\text{NO}_3^-]) (µmol kg(^{-1}))</th>
<th>([\text{NO}_2^-]) (µmol kg(^{-1}))</th>
<th>([\text{NH}_4^+]) (µmol kg(^{-1}))</th>
<th>([\text{PO}_4^{3-}]) (µmol kg(^{-1}))</th>
<th>SD</th>
<th>([\text{NO}_3^-]) (µmol kg(^{-1}))</th>
<th>([\text{NO}_2^-]) (µmol kg(^{-1}))</th>
<th>([\text{NH}_4^+]) (µmol kg(^{-1}))</th>
<th>([\text{PO}_4^{3-}]) (µmol kg(^{-1}))</th>
<th>SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.16</td>
<td>0.17</td>
<td>0.03</td>
<td>0.02</td>
<td>0.25</td>
<td>0.11</td>
<td>0.01</td>
<td>0.01</td>
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<tr>
<td>20</td>
<td>0.21</td>
<td>0.20</td>
<td>0.01</td>
<td>0.01</td>
<td>0.20</td>
<td>0.07</td>
<td>0.01</td>
<td>0.01</td>
<td>1.78</td>
<td>1.40</td>
</tr>
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<td>0.35</td>
<td>0.25</td>
<td>0.02</td>
<td>0.02</td>
<td>0.34</td>
<td>0.17</td>
<td>0.02</td>
<td>0.00</td>
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<td>2.02</td>
</tr>
<tr>
<td>100</td>
<td>4.66</td>
<td>0.69</td>
<td>0.15</td>
<td>0.07</td>
<td>0.17</td>
<td>0.06</td>
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<td>0.04</td>
<td>8.09</td>
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</tr>
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<td>0.04</td>
<td>0.03</td>
<td>0.04</td>
<td>10.38</td>
<td>0.90</td>
<td>1.03</td>
<td></td>
</tr>
<tr>
<td>250</td>
<td>4.28</td>
<td>3.98</td>
<td>1.08</td>
<td>0.51</td>
<td>3.71</td>
<td>2.02</td>
<td>1.32</td>
<td>0.44</td>
<td>3.71</td>
<td>2.02</td>
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<td>0.16</td>
<td>3.66</td>
<td>1.28</td>
<td>2.91</td>
<td>1.51</td>
<td>0.30</td>
<td>0.33</td>
<td>2.91</td>
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<td>400</td>
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<td>0.13</td>
<td>0.05</td>
<td>7.81</td>
<td>2.48</td>
</tr>
</tbody>
</table>

Fig. 3. Salinity versus potential temperature (\(\theta\)) in the Cariaco Basin at stations sampled for \(N_{2}^{ss}\) measurements during September 2008 and March 2009. Black symbols represent the average potential temperature and salinity from all six stations and sampling periods at 150 (diamond), 200 (square) and 250 (circle) meters.

4.3 \(N_2/Ar\) ratios and \(N_{2}^{ss}\)

Average vertical distributions of normalized \(N_2/Ar\) ratios during both cruises are shown in Fig. 5a. Average normalized \(N_2/Ar\) ratios measured in the upper 100 m were undersaturated by \(\sim 0.5\%\) in September 2008 and \(\sim 0.2\%\) in March 2009. The mean \(N_2/Ar\) ratio near the oxic–anoxic interface (250–300 m) was 1.026 (\(\sim 2.6\%\) supersaturated) during both periods. Below this depth, \(N_2/Ar\) ratios remained fairly constant. Estimates of background \(N_2/Ar\) ratios (calculated based on regressions of the ratio versus temperature reported at BATTS) in surface waters were close to atmospheric equilibrium in September 2008 and \(\sim 0.3\%\) supersaturated in March 2009. Corresponding background \(N_{2}^{ss}\) in the upper 100 m showed negative values in September 2008 and positive values in March 2009 (\(\sim 2\mu\text{mol N kg}^{-1}\); Fig. 5b). Estimated background \(N_2/Ar\) ratios peaked at 200 m (\(\sim 0.6\%\) supersaturation) with background \(N_{2}^{ss}\) at \(\sim 6\mu\text{mol N kg}^{-1}\) during both periods and remaining constant below this depth (Fig. 5a, b).

In March 2009, observed \(N_{2}^{ss}\) in surface waters was closer to equilibrium with the atmosphere. In September 2008, waters in the upper 100 m had observed \(N_{2}^{ss}\) negative anomalies as high as \(\sim 8\mu\text{mol N kg}^{-1}\), presumably in response to stratification and warming by 4°C as compared to March 2009. Waters \(\geq 200\) m were supersaturated with dissolved \(N_2\) at all stations during both cruises. Below the upper 150 m, observed \(N_{2}^{ss}\) rapidly increased to peak concentrations near the oxic–anoxic interface (250–300 m). In March 2009, \([N_{2}^{ss}]\) decreased by \(\sim 3\mu\text{mol N kg}^{-1}\) from these maximal values at depths below 400 m. In addition, samples collected between 300–400 m at four stations during the upwelling period (March 2009) had significantly higher \([N_{2}^{ss}]\) (2.7–6.1 µmol N kg\(^{-1}\), \(p < 0.001\)) than those at the same stations during the relaxation period (September 2008) (Fig. 6). This was not observed at stations 10 (in the center of the eastern basin) and 29 (in the western basin near the Tuy River). However, we did not detect a clear spatial pattern along sampled stations which would have suggested an influence from riverine discharge or Caribbean waters entering into the basin through the Centinela and Tortuga channels (Fig. 1) on \(N_{2}^{ss}\) distributions.

5 Discussion

5.1 \(N_2/Ar\) ratios and \(N_{2}^{ss}\)

Surface waters of the Cariaco Basin showed negative \(N_{2}^{ss}\) anomalies during both periods (Fig. 5a and 5b). The magnitude of \(N_{2}^{ss}\) negative anomalies was lower in March 2009 than in September 2008, when sea surface temperatures
Fig. 4. Nutrient profiles from the CARIACO Time Series Station during September 2008 and March 2009.

Fig. 5. Profiles of normalized $\text{N}_2/\text{Ar}$ ratios and $[\text{N}_2^{\text{SS}}]$ in the Cariaco Basin during September 2008 and March 2009. Background (abiotic) $\text{N}_2/\text{Ar}$ ratios and $[\text{N}_2^{\text{SS}}]$ were estimated using data collected at BATS (see Sect. 3.2).
(SST) were significantly higher (\(\sim 29^\circ\text{C}\)). Hence, in situ near-surface water heating without gas re-equilibration is the most likely cause for the negative values of \(\Delta [N_2]^{\text{S}}\). Increasing SST reduces the equilibrium concentration of \(N_2\) to a lesser degree than Ar, thereby increasing equilibrium \(N_2/\text{Ar}\). If actual \(N_2\) and Ar concentrations have little to no change due to low air–sea gas exchange, then the observed \(N_2/\text{Ar}\) is less than the equilibrium value and a negative \([\Delta N_2]\) results. Calculations (not shown) show that the magnitude of observed negative \([\Delta N_2]\) is consistent with the seasonal temperature increase assuming in situ heating with minimal gas exchange.

\([\Delta N_2]\) values were relatively constant with depth in the upper 100 m during both sampling periods but increased below 200 m, as dissolved oxygen decreased and N-loss began. The \([\Delta NO_3]\) profile shows the consumption of nitrate by denitrification between \(\sim 200\) and 300 m (Fig. 4). In most stations, the highest \([\Delta NO_3]\) \((\sim 12–30\ \mu\text{mol N kg}^{-1})\;\text{avg} = 23.8 \pm 4.3\ \mu\text{mol N kg}^{-1}\) was measured at 250–300 m (Fig. 5). The mean \([\Delta N_2]\) between the oxic–anoxic interface (250 m) and 1300 m depth was \(25 \pm 2\ \mu\text{mol N kg}^{-1}\), which is equivalent within uncertainty to the increase in \(N_2\) expected from the N deficit \((\sim 23\ \mu\text{mol N kg}^{-1})\). Our primary assumption is that the basin is filled with silt-depth \((\sim 150\text{ m})\) Caribbean Sea intermediate waters that were once fully oxygenated, as previously proposed by Richard and Benson (1961). In the Cariaco Basin nitrogen derives from two sources: incoming nitrate and N from remineralized organic matter. Unpublished data show that nitrate and oxygen concentrations in Caribbean intermediate waters entering the basin are 6–8 and \(\sim 140\ \mu\text{mol kg}^{-1}\), respectively (L. Lorenzoni unpublished data, 2004). Subsequent lack of ventilation induces today’s anoxic conditions and thus denitrification leading to an N deficit and buildup of \(N_2\) above pre-existing background levels. The consumption of nitrate by denitrification would thus have generated \(\sim 7\ \mu\text{mol N kg}^{-1}\) of \(N_2\), while the consumption of oxygen by organic matter remineralization would have produced \(\sim 16\ \mu\text{mol kg}^{-1}\) of DIN available for N-loss (AOU : N = \(\sim 270 : 16\) for an expected total of \(\sim 23\ \mu\text{mol N kg}^{-1}\) of \(N_2\) produced. Oxygen consumption by organic matter remineralization would have released ammonium \((\sim 16\ \mu\text{mol kg}^{-1})\) that would have subsequently been either directly consumed by anammox or transformed into \(\text{NO}_3\) available for denitrification. The entire process is likely very dynamic and the observed \(N_2\) in the Cariaco Basin is the result of multiple pulses of Caribbean Sea intermediate waters flowing over the sill into the suboxic and anoxic portions of the basin. Evidence of such intrusion events have been reported by Astor et al. (2003) and Scranton et al. (2006). Nitrate from intrusions into the oxic–anoxic interface would be consumed by denitrification while that injected into the sulfidic portion of the water column would be consumed by autotrophic denitrification, a biological process that uses sulfur species as reductant to convert nitrate into \(N_2\) (Fuchsman et al., 2012; Jensen et al., 2009). Though an overlap between sulfide and nitrate has not been detected in the basin (M. Scranton, personal communication, 2012) and incubations have not shown any stimulation of dark carbon fixation by nitrate, previous work has reported high rates of dark dissolved inorganic carbon (DIC) fixation (chemoautotrophy) occurring at the oxic–anoxic interface in the Cariaco Basin (Taylor et al., 2001, 2006; Wakeham et al., 2012).

Our profiles of \(N_2/\text{Ar}\) ratios and \([\Delta N_2]\) are similar in shape to the earlier profiles measured by Richards and Benson (1961) in the Cariaco Basin. They found an average \([\Delta N_2]\) of \(20.7 \pm 2.1\ \mu\text{mol kg}^{-1}\) at \(\sim 400–1200\) m, or \(\sim 25\%\) lower than our average \([\Delta N_2]\) of \(25.0 \pm 1.8\ \mu\text{mol N kg}^{-1}\) in the same depth range in the eastern sub-basin (stations 10, 11 and 13). It is unlikely that such difference in \([\Delta N_2]\) results from methodological discrepancies. Richards and Benson’s \(N_2\) measurements were carried out following the methods described in Benson and Parker (1961). Differences between their methodology and ours included sample storage in specially made piston sealed bottles and dissolved gas extraction on a vacuum line prior to transfer to a mass spectrometer using a Toepler pump. They also removed \(O_2\) using hot copper prior to \(N_2/\text{Ar}\) analysis. While the method used by Richards and Benson (1961) is more time-consuming than ours, there is no reason to suspect a systematic bias between our approaches as they also determined that they achieved quantitative gas extraction. In fact, Richards and Benson (1961) also observed slight negative \([\Delta N_2]\) of 1 to 3 \(\mu\text{mol N kg}^{-1}\) as we do. Such agreement would be unlikely if either of our methods has significant bias. To our knowledge, these are the only two studies of \(N_2\) that have been conducted in the Cariaco Basin. The difference suggests that \(N_2\) has been accumulating in the basin at a rate of \(\sim 0.08\ \mu\text{mol N kg}^{-1}\ yr^{-1}\) (from 1957 to 2009) within this depth interval \((\sim 400–1200\) m). We see a concurrent increase in SRP, ammonium and silicate concentrations (0.035–0.037, 0.28–0.62 µmol kg\(^{-1}\)).
and 0.85–1.3 µmol kg\(^{-1}\) yr\(^{-1}\), respectively; Scranton et al., 2006; Zhang and Millero, 1993) The increase in SRP better matches Redfield stoichiometry (15–16: 1 N : P) when both the accumulation rate of \(N_2^{\text{ox}}\) (0.08 µmol N kg\(^{-1}\) yr\(^{-1}\)) and ammonium (0.45 µmol N kg\(^{-1}\) yr\(^{-1}\); average based on values reported above) are accounted for. The DIN/SRP ratio of the accumulation is 14.7 (0.08 + 0.45 : 0.036) if \(N_2^{\text{ox}}\) is included and 12.5 when it is not.

5.2 Biogenic \(N_2\) production and upwelling

The \([N_2^{\text{ox}}]\) measured immediately below the oxic–anoxic interface at 300–400 m in March 2009 (upwelling) is higher than that measured in September 2008 \((p < 0.001)\) (non-upwelling; Fig. 6), suggesting that N-loss and thus production of biogenic \(N_2\) are stimulated during upwelling in the Cariaco Basin. The upwelling period usually is associated with higher fluxes of settling particulate organic matter (Montes et al., 2012; Thunell et al., 2007). Enhanced vertical transport of particulate organic matter can be an important mode of supply of fixed nitrogen to the oxic–anoxic interface, where degradation of the flux releases ammonium and organic carbon that can be utilized by microbes. This is consistent with findings by Fuchsun et al. (2008) in the Black Sea, who observed higher production of biogenic \(N_2\) in years that had large phytoplankton blooms compared to those in which phytoplankton abundances were lower. Nutrient and suspended organic particle measurements from the CARIACO Ocean Time Series program show that DIN, POC and particulate organic nitrogen (PON) at the oxic–anoxic interface \((\sim 250\ m)\) were higher in March 2009 than in September 2008 (5.18 vs. 1.02 µmol N kg\(^{-1}\), 65.12 vs. 46.04 µg C kg\(^{-1}\) and 11.16 vs. 8.05 µmol N kg\(^{-1}\); respectively). Furthermore, particulate settling flux records from CARIACO show that mean PON fluxes \((1997–2010)\) at \(\sim 400\ m\) tend to be higher in March than in September months (Fig. 7); mean PON fluxes for CARIACO were used since settling flux data were not available for 2009 because of a mooring malfunction. These observations suggest that more inorganic and organic forms of dissolved nitrogen are available during upwelling compared to non-upwelling periods, stimulating N-loss processes during upwelling.

Ammonium produced by the decay of sinking organic matter, in particular, is likely an important DIN source for N-loss. It can be incorporated into non-cannotial nitrogen pathways such as coupled nitrification–denitrification in which nitrifiers exploit the newly generated ammonium at the oxic–anoxic interface, producing nitrite and nitrate that can, in turn, be utilized by heterotrophic denitrifiers. Another plausible utilization pathway of \(\text{NO}_3^-\) is anammox. Historical nutrient records from the CARIACO Ocean Time Series program indicate that \(\text{NO}_3^-\) is occasionally present at the interface in high enough concentrations to fuel the anammox reaction (Scranton et al., 2006). Wakeham et al. (2012) also shows extensive evidence for anammox taking place at the oxic–anoxic interface in Cariaco. They found that anammox bacteria and ladinene lipids, an anammox bacterial biomarker, were abundant at \(\sim 250\ m\). Furthermore, maxima in chemotrophic production rates (dark DIC assimilation) have also been measured at this depth, likely as a result of the presence of the anammox metabolism and perhaps autotrophic denitrification (or thiodenitrification) within this layer (Taylor et al., 2001, 2006; Wakeham et al., 2012).

From the observations reported in this study, however, we cannot determine which specific pathways (denitrification or anammox) dominate the conversion of DIN to biogenic \(N_2\) within Cariaco’s sub-oxic layer. Nonetheless, from a general perspective our results shed light on the microbiologically mediated biogeochemical processes that impact the production of biogenic \(N_2\) in the basin on a seasonal basis. Here we propose a model explaining the response of the N-loss to the upwelling cycle of Cariaco. The rain of organic matter is an important source of carbon and nitrogen for autotrophs and heterotrophs at the oxic–anoxic interface. Previous work has shown that seasonal changes in bacterial respiration rates within this layer can be associated with changes in the upwelling conditions in the basin (Taylor et al., 2006, 2009). Our hypothesis is that enhanced settling flux of organic matter during the upwelling period stimulates both heterotrophic and autotrophic metabolism at the oxic–anoxic interface. Though in a different setting, Lomas et al. (2010) found that Sargasso Sea’s mesopelagic export production \((300\ m)\) has remained constant since the late 90’s, while epipelagic export

Fig. 7. Mean vertical PON flux during September (non-upwelling) and March (upwelling) months from the CARIACO Ocean Time Series between 1997 and 2010.
Particulate P has nanomolar concentrations at this depth appear to be minimally impacted (Montes et al., 2012), a decrease of the microbial activity can be expected, which would lead to lower production of biogenic N$_2$.

An alternative explanation for the increase in biogenic N$_2$ is water mass intrusions at intermediate depths. Episodic intrusions of oxygen and nutrient-rich water masses from the Caribbean Sea deepen the oxic–anoxic interface in Cariaco (Astor et al., 2003). These alter the vertical distribution of redox sensitive species such as hydrogen sulﬁde, iron and manganese oxides, and ammonium, among others (Percy et al., 2008; Scranton et al., 2001, 2006). Dissolved oxygen concentrations were higher between the ~25 and ~26 isopycnal surfaces in March 2009 compared to September 2008 (Fig. 2). The increase in dissolved O$_2$ during the upwelling season must result from enhanced vertical mixing and ventilation of cold (~21–22°C) subsurface waters. Higher [O$_2$] in the upper ~120 m in March 2009 suggests that an intrusion of water from the Caribbean Sea could have occurred during the upwelling period. The supply of oxygen to the redoxcline would stimulate nitrification, and enhance the abundance of NO$_x$ available for denitriﬁers. Though an increase in oxygen concentrations at this depth after an intrusion would inhibit denitriﬁcation, previous work has shown that oxygen is quickly consumed and that low oxygen conditions are restored within weeks at this depth (Scranton et al., 2006). The intrusion of nitrate-rich waters from the Caribbean would also stimulate denitriﬁcation at the oxic–anoxic interface.

5.3 DIN deﬁcit and N$^*_2$

We estimate the DIN deﬁcit ($N_{\text{def}}$) in the basin as the amount of ﬁxed nitrogen that has been removed by the N-loss (Chang et al., 2010; Devol et al., 2006). The $N_{\text{def}}$ is calculated by subtracting the observed DIN ($N_{\text{obs}}$) from the total expected fixed nitrogen ($N_{\text{exp}} = [\text{NO}_3^-]+ [\text{NO}_2^-]+ [\text{NH}_4^+]$) based on in situ SRP concentrations and Redﬁeld stoichiometry. DIN deﬁcits within the anoxic portion of the water column in Cariaco are not likely affected by reduced metals adsorption. Unpublished data show that despite labile and oxide-associated particulate P peak at Cariaco’s oxic–anoxic interface, indicating SRP scavenging by reduced metals, SRP concentrations at this depth appear to be minimally impacted by such reactions (C. Benitez-Nelson, personal communication, 2012). Particulate P has nanomolar concentrations while SRP concentrations are in the micromolar range. The expected DIN abundance is typically obtained by plotting DIN versus SRP concentrations measured in waters with dissolved O$_2 > 65$ µmol kg$^{-1}$ (Devol et al., 2006). For our work we used nutrient data collected at the CARIACO Station and BATS during 2008 and 2009 using the same criteria as Devol et al. (Fig. 8). Data from the BATS program was used in this analysis since waters entering into the Cariaco Basin originate in the subtropical North Atlantic. $N_{\text{exp}}$ is then calculated by linear regression:

\[ N_{\text{exp}} = 14.5(P_{\text{obs}}) - 0.7, \]  

(1)

where $P_{\text{obs}}$ corresponds to in situ SRP concentrations. The negative Y-intercept indicates a DIN deﬁcit when the SRP concentration is zero. The slope and intercept of 14.5 and 0.7, respectively, are the result of averaging the corresponding values from the DIN : SRP relationships from the CARIACO Time Series (13.8, 0.5) and BATS (15.1, 0.9). These N : P relationships are similar to those reported from the eastern tropical South Paciﬁc (15.8) and the Arabian Sea (14.9) (Chang et al., 2010, and Codispoti et al., 2001, respectively). The DIN deﬁcit is then

\[ N_{\text{def}} = (N_{\text{exp}} - N_{\text{obs}}). \]  

(2)

We based $N_{\text{def}}$ estimates on monthly observations at the CARIACO Station (station 10 in this study) for 2008–2009 since no nitrite or SRP measurements were made at depths below 100 m during our cruises. We believe that these are representative for the basin because the vertical nutrient proﬁles within each season were similar among all stations during the regional cruises at the oxic–anoxic interface (Table 2).

DIN deﬁcits in September 2008 and March 2009 were calculated by averaging the $N_{\text{def}}$ for the CARIACO observations between August and October 2008, and between February and April 2009, respectively. Combined $N_{\text{def}}$ in the upper ~120 m were below detection during both seasons, as expected, but increased in waters with low oxygen (≥ 200 m). Measurements of [N$^*_2$] were similar to $N_{\text{def}}$ (least squares linear correlation, $R^2 = 0.95$, $p < 0.001$; Fig. 9), showing that biogenic N$_2$ production reﬂects the amount of fixed nitrogen removed by the N-loss process in Cariaco. Biogenic [N$^*_2$] tended to be slightly lower than $N_{\text{def}}$ estimates (Fig. 9), likely due to the accumulation of SRP within the anoxic layer, which artiﬁcially ampliﬁes $N_{\text{def}}$ values.

These observations support the conclusion of Chang et al. (2010) that measurements of N$^*_2$ provide a useful means for estimating N-loss rates in oxygen minimum zones, and that N$^*_2$ concentrations can be used to validate N-loss rates in marine systems (see DeVries et al., 2012) where DIN deﬁcits or N$^*_2$ estimates are affected by non-Redﬁeld N : P ratios. Some potential problems of using DIN deﬁcits calculated from SRP include variable N : P ratios, the scavenging of SRP by redox species such as iron and manganese oxides, or (speciﬁcally for Cariaco) the accumulation of SRP.
and ammonium below the oxic–anoxic interface (Scranton et al., 2006; Zhang and Millero, 1993). The excellent agreement between [N$_{xs}^2$] and N$_{def}$ supports our assertion that N$_{xs}^2$ in the Cariaco Basin is mainly of biological origin.

6 Conclusions

In this study we evaluated changes in N$_{xs}^2$ in the Cariaco Basin under contrasting seasonal conditions (upwelling versus non-upwelling). [N$_{xs}^2$] were below detection at the surface, and increased to a mean value of $\sim$ 22 $\mu$mol N kg$^{-1}$ at the oxic–anoxic interface (250–300 m) and deeper. Our [N$_{xs}^2$] estimates are $\sim$ 25 % larger than those from Richards and Benson (1961), suggesting that biogenic N$_2$ has been accumulating in the basin at a rate of $\sim$ 0.08 $\mu$mol N kg$^{-1}$ yr$^{-1}$.

[N$_{xs}^2$] was higher by $\sim$ 3–6 $\mu$mol N kg$^{-1}$ at 300–400 m during the upwelling period (March 2009) than when the basin was more thermally stratified (September 2008). Higher biogenic N$_2$ production can be the result of enhanced DIN availability in Cariaco’s oxic–anoxic interface during the upwelling period due to higher vertical transport of organic matter.

N-loss at the oxic–anoxic interface of the Cariaco Basin appears to be stimulated during the upwelling period. We attribute this observation to two mechanisms: (1) increased supply of sinking organic nitrogen and/or (2) ventilation of the oxic–anoxic interface by intermediate oxygenated and nitrate-rich Caribbean waters.

DIN deficits based on nutrient Redfield stoichiometry within sub-oxic and anoxic waters in Cariaco are similar to our estimates of [N$_{xs}^2$] at the corresponding depths. This suggests that N$_{xs}^2$ in Cariaco is generated mainly by biological processes associated with oceanic N-loss.

Our observations show that measuring N$_{xs}^2$ is a robust method for quantifying the sink of fixed nitrogen in Cariaco and in low oxygen environments in general. We conclude that biogenic N$_2$ determinations are a useful tool for complementing information provided by the N* parameter, and should be considered in future studies addressing questions regarding the state of the contemporary global nitrogen cycle.

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